

Disease Classification

OMB No: 0915-0310 Expiration Date: 08/31/2025

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CIBMTR Center Number:		
CIBMTR Research ID:		
Event date:	-	
YYYY MM DD		

CIBM	ITR C	enter Number: CIBMTR Research ID:
Prim	ary D	isease for HCT / Cellular Therapy
1.	Date	e of diagnosis of primary disease for HCT / cellular therapy:
2.	Wha	at was the primary disease for which the HCT / cellular therapy was performed?
		Acute myeloid leukemia (AML) (10) – Go to question 3
		Acute lymphoblastic leukemia (ALL) (20) – Go to question 104
		Acute leukemia of ambiguous lineage and other myeloid neoplasms (80) – <i>Go to question 180</i>
		Chronic myeloid leukemia (CML) (40) – Go to question 184
		Myelodysplastic syndrome (MDS) (50) (If recipient has transformed to AML, indicate AML as the primary disease.) – Go to question 195
		Myeloproliferative neoplasms (MPN) (1460) (If recipient has transformed to AML, indicate AML as the primary disease.) – Go to question 275
		Other leukemia (30) (includes CLL) - Go to question 388
		Hodgkin lymphoma (150) – <i>Go to question 395</i>
		Non-Hodgkin lymphoma (100) – <i>Go to question 395</i>
		Multiple myeloma / plasma cell disorder (PCD) (170) – Go to question 413
		Solid tumors (200) – Go to question 460
		Aplastic anemia (300) (If the recipient developed MDS or AML, indicate MDS or AML as the primary disease.) – Go to question 462
		Inherited bone marrow failure syndromes (320) (If the recipient developed MDS or AML, indicate MDS or AML as the primary disease.) – Go to question 465
		Hemoglobinopathies (330) – Go to question 466
		Paroxysmal nocturnal hemoglobinuria (PNH) (340) – Go to end of form
		Disorders of the immune system (400) – Go to question 502
		Inherited abnormalities of platelets (500) – Go to question 510
		Inherited disorders of metabolism (520) – <i>Go to question 512</i>
		Histiocytic disorders (570) – <i>Go to question 515</i>
		Autoimmune diseases (600) – Go to question 520
		Tolerance induction associated with solid organ transplant (910) - Go to question 524
		Recessive dystrophic epidermolysis bullosa (920) – Go to end of form
		Other disease (900) – Go to question 526

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Acute Myeloid Leukemia (AML)

4.

3. Specify the AML classification

AM	L with defining genetic abnormalities Acute myeloid leukemia with MLLT3::KMT2A fusion (5)
	Acute myeloid leukemia with other KMT2A rearrangements (284)
	Acute myeloid leukemia with DEK::NUP214 fusion (6)
	Acute myeloid leukemia with MECOM (EVI1), GATA2 rearrangement (7
	Acute myeloid leukemia with Other MECOM rearrangements (1011)
	Acute myeloid leukemia with RBM15::MRTFA fusion (8)
	Acute myeloid leukemia with RUNX1::RUNX1T1 fusion (281)
	Acute myeloid leukemia with CBFB::MYH11 fusion (282)
	Acute promyelocytic leukemia with PML::RARA fusion (283)
	Acute promyelocytic leukemia with other RARA fusions (1012)
	Acute myeloid leukemia with BCR::ABL1 fusion (3)
	Acute myeloid leukemia with NPM1 mutation (4)
	Acute myeloid leukemia with CEBPA mutation (297)
	Acute myeloid leukemia with myelodysplasia – related (285)
	Acute myeloid leukemia with NUP98 rearrangement (1013)
	Acute myeloid leukemia with mutated TP53 (1014)
	Acute myeloid leukemia with other defined genetic alterations (1015)
AM	L, defined by differentiation Acute myeloid leukemia with minimal differentiation (286)
	Acute myeloid leukemia without maturation (287)
	Acute myeloid leukemia with maturation (288)
	Acute myelomonocytic leukemia (289)
	Acute monocytic leukemia (290)
	Acute erythroid leukemia (291)
	Acute megakaryoblastic leukemia (292)
	Acute basophilic leukemia (293)
	Myeloid sarcoma (295)
	Acute myeloid leukemia, not otherwise specified (280)
Did	AML transform from MDS or MPN?
	Yes – Also complete MDS or MPN Disease Classification questions
	No

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5.	Is the disease (AML) therapy related?
	□ Yes
	□ No
	□ Unknown
6.	Did the recipient have a predisposing condition?
	☐ Yes – Go to question 7
	□ No – Go to question 9
	□ Unknown – Go to question 9
	7. Specify condition
	☐ Bloom syndrome – <i>Go to question 9</i>
	☐ Down syndrome – Go to question 9
	☐ Fanconi anemia – Also complete CIBMTR Form 2029 – FAN – Go to question 9
	☐ Dyskeratosis congenita – Also complete CIBMTR Form 2028 – APL – Go to question 9
	☐ Other condition – Go to question 8
	8. Specify other condition:
	Laboratory studies at diagnosis
9.	Were cytogenetics tested (karyotyping or FISH)? (at diagnosis)
	☐ Yes – Go to question 10
	□ No – Go to question 23
	☐ Unknown – Go to question 23
	10. Were cytogenetics tested via FISH?
	☐ Yes – Go to question 11
	□ No – Go to question 16
	11. Results of tests
	☐ Abnormalities identified – <i>Go to question 12</i>
	☐ No abnormalities – <i>Go to question 16</i>
	12. International System for Human Cytogenetic Nomenclature (ISCN) compatible strin
	13. Specify number of distinct cytogenetic abnormalities□ One (1)
	□ Two (2)

CIBMTR Center Number:	CIBMTR Research ID:
	Three (3)
	Four or more (4 or more)
14. Spe	cify abnormalities <i>(check all that apply)</i>
·	-5
	-7
	-17
	-18
	-X
	-Y
	+4
	+8
	+11
	+13
	+14
	+21
	+22
	t(3;3)
	t(6;9)
	t(8;21)
	t(9;11)
	t(9;22)
	t(15;17) and variants
	t(16;16)
	del(3q) / 3q-
	del(5q) / 5q-
	del(7q) / 7q-
	del(9q) / 9q-
	del(11q) / 11q-
	del(16q) / 16q– del(17q) / 17q–
	del(20q) / 20q-
	del(21q) / 21q-
	inv(3)
	inv(16)
_	(11q23) any abnormality

☐ 12p any abnormality

CIBMTR Center Numbe	er:		CIBMTR Research ID:
			Other abnormality – <i>Go to question 15</i>
	1	15.	Specify other abnormality:
16. We	ere cytogene	etics	tested via karyotyping?
	Yes – Go t	o q	uestion 17
	No – Go to	qu	estion 22
17	'. Results	of te	ests
	☐ Abno	orma	alities identified – Go to question 18
	□ No e	valu	able metaphases – <i>Go to question 22</i>
	□ No a	bno	rmalities – Go to question 22
	18. I	nter	national System for Human Cytogenetic Nomenclature (ISCN) compatible string:
	19.	Spe	cify number of distinct cytogenetic abnormalities
			One (1)
			Two (2)
			Three (3)
			Four or more (4 or more)
	20.	Spe	cify abnormalities (check all that apply)
			-5
			-7
			-17
			-18
			-X
			-Y
			+4
			+8
			+11
			+13
			+14
			+21
			+22
			t(3;3)
			t(6;9)
			t(8;21)

CIBMTR Center Number:	CIBMTR Research ID:
	□ t(9;11)
	□ t(9;22)
	□ t(15;17) and variants
	□ t(16;16)
	□ del(3q) / 3q-
	□ del(5q) / 5q-
	□ del(7q) / 7q-
	□ del(9q) / 9q-
	□ del(11q) / 11q–
	□ del(16q) / 16q-
	□ del(17q) / 17q–
	□ del(20q) / 20q-
	□ del(21q) / 21q-
	□ inv(3)
	□ inv(16)
	□ (11q23) any abnormality
	□ 12p any abnormality
	☐ Other abnormality – <i>Go to question 21</i>
	21. Specify other abnormality:
22. Was docum	entation submitted to the CIBMTR? (e.g. cytogenetic or FISH report)
☐ Yes	
□ No	
23. Were tests for mole	cular markers performed? (e.g. PCR, NGS) (at diagnosis)
☐ Yes – Go to qu	uestion 24
□ No – Go to que	estion 36
□ Unknown – <i>Go</i>	to question 36
24. CEBPA	
☐ Positive -	- Go to question 25
☐ Negative	– Go to question 26
☐ Not done	- Go to question 26
25. Speci	fy CEBPA mutation
□ Bia	allelic (double mutant)
□Мо	onoallelic (single mutant)

CIBMTR Center Nu	mber: CIBMTR Research ID:
	□ Unknown
26.	FLT3 – TKD (point mutations in D835 or deletions of codon I836)
	□ Positive
	□ Negative
	□ Not done
27.	FLT3 – ITD mutation
	□ Positive – Go to question 28
	□ Negative – Go to question 30
	□ Not done – Go to question 30
	28. FLT3 – ITD allelic ratio
	☐ Known – Go to question 29
	☐ Unknown – Go to question 30
	29. Specify FLT3 - ITD allelic ratio:
30.	IDH1
	□ Positive
	□ Negative
	□ Not done
31.	IDH2
	□ Positive
	□ Negative
	□ Not done
32.	KIT
	□ Positive
	□ Negative
	□ Not done
33.	NPM1
	□ Positive
	□ Negative
	□ Not done

Copy and complete questions 34-35 for multiple molecular markers

CIBMTR Center Number: _	CIE	BMTR Research ID:
34. Other r	nolecular marker	
□ Pos	itive – Go to question 35	
□ Neg	ative – Go to question 3	5
□ Not	done - Go to question 3	5
35.	Specify other molecular ma	rker:
Copy and complete	questions 34-35 for multi	ple molecular markers
Labs between diagrathe last evaluation t		(If subsequent infusion, labs after prior infusion and before
36. Were cytogene	etics tested (karyotyping or	FISH)? (in between)
□ Yes – Go	to question 37	
□ No – Go t	o question 50	
□ Unknown	– Go to question 50	
37. Were o	ytogenetics tested via FISH	I?
☐ Yes	– Go to question 38	
□ No	- Go to question 43	
38. I	Results of tests	
	☐ Abnormalities identified -	- Go to question 39
	□ No abnormalities – Go t	·
		•
	39. International System	n for Human Cytogenetic Nomenclature (ISCN) compatible string:
	40. Specify number of d	istinct cytogenetic abnormalities
	□ One (1)	
	□ Two (2)	
	☐ Three (3)	
	☐ Four or more (4 or more)
	41. Specify abnormalitie	s (check all that apply)
	□ -5	
	□ -7	
	□ -17	
	□ -18	
	□ -X	

CIBMTR Center Number:	CIBMTR Research ID:
	-Y
	+4
	+8
	+11
	+13
	+14
	+21
	+22
	t(3;3)
	t(6;9)
	t(8;21)
	t(9;11)
	t(9;22)
	t(15;17) and variants
	t(16;16)
	del(3q) / 3q-
	del(5q) / 5q-
	del(7q) / 7q-
	del(9q) / 9q-
	del(11q) / 11q-
	del(16q) / 16q-
	del(17q) / 17q-
	del(20q) / 20q-
	del(21q) / 21q-
	inv(3)
	inv(16)
	(11q23) any abnormality
	12p any abnormality
	Other abnormality – Go to question 42
42.	Specify other abnormality:
43. Were cytogenetics	s tested via karyotyping?
☐ Yes – Go to q	uestion 44
□ No – Go to qu	restion 49
44. Results of to	ests

CIBMTR Center Number:			CIBMTR Research ID:	
	☐ No evaluable metaphases – <i>Go to question 49</i>			
	□ No	abno	rmalities – Go to question 49	
	45. Inter		rnational System for Human Cytogenetic Nomenclature (ISCN) compatible string:	
	46.	Spe	cify number of distinct cytogenetic abnormalities	
			One (1)	
			Two (2)	
			Three (3)	
			Four or more (4 or more)	
	47.	Spe	cify abnormalities (check all that apply)	
			-5	
			-7	
			-17	
			-18	
			-X	
			-Y	
			+4	
			+8	
			+11	
			+13	
			+14	
			+21	
			+22	
			t(3;3)	
			t(6;9)	
			t(8;21)	
			t(9;11)	
			t(9;22)	
			t(15;17) and variants	
			t(16;16)	
			del(3q) / 3q-	
			del(5q) / 5q-	
			del(7q) / 7q-	
			del(9q) / 9q-	
			del(11q) / 11q-	

CIBMTR Center Number:	CIBMTR Research ID:		
	del(16q) / 16q–		
	del(17q) / 17q-		
	del(20q) / 20q-		
	del(21q) / 21q-		
	inv(3)		
	inv(16)		
	(11q23) any abnormality		
	12p any abnormality		
	Other abnormality – <i>Go to question 48</i>		
48.	Specify other abnormality:		
49. Was documentation	on submitted to the CIBMTR? (e.g. cytogenetic or FISH report)		
☐ Yes			
□ No			
50. Were tests for molecular	markers performed? (e.g. PCR, NGS) (in between)		
☐ Yes – Go to question	on 51		
□ No – Go to question	n 63		
☐ Unknown – Go to qu	uestion 63		
51. CEBPA			
☐ Positive – Go	to question 52		
☐ Negative – Go	to question 53		
☐ Not done – Go	to question 53		
52. Specify CEI	BPA mutation		
•	(double mutant)		
☐ Monoalle	elic (single mutant)		
□ Unknow	n		
53. FLT3 – TKD <i>(poin</i>	t mutations in D835 or deletions of codon I836)		
☐ Positive			
☐ Negative			
☐ Not done			
54. FLT3 – ITD mutati	ion		
☐ Positive – <i>Go</i> a	to question 55		
□ Negative – <i>Go</i>	to question 57		

CIBMTR Center Nu	mber: CIBMTR Research ID:					
	□ Not done – <i>Go to question 57</i>					
	55. FLT3 – ITD allelic ratio					
	☐ Known – <i>Go to question 56</i>					
	☐ Unknown – Go to question 57					
	56. Specify FLT3 - ITD allelic ratio:					
57.	IDH1					
	□ Positive					
	□ Negative					
	□ Not done					
58.	IDH2					
	□ Positive					
	□ Negative					
	□ Not done					
59.	KIT					
	□ Positive					
	□ Negative					
	□ Not done					
60.	NPM1					
	□ Positive					
	□ Negative					
	□ Not done					
Сору	and complete questions 61-62 to report multiple other molecular markers					
61.	Other molecular marker:					
	□ Positive – Go to question 62					
	□ Negative – Go to question 62					
	□ Not done – Go to question 63					
	62. Specify other molecular marker:					

Copy and complete questions 61-62 to report multiple other molecular markers

Labs at last evaluation

CIBMTR Center Number:				CIBMTR Research ID:				
	63.	63. Were cytogenetics tested (karyotypi			ested	(karyotyping or FISH)? (at last evaluation)		
			☐ Yes – Go to question 64					
			No – Go to question 77					
			Unknowr	n – Go	to qu	restion 77		
		64	. Were	cvtoge	netics	s tested via FISH?		
				-		uestion 65		
					_	estion 70		
			65.	Result	s of te	ests		
			00.		onormalities identified – <i>Go to question 66</i>			
						rmalities – <i>Go to question 70</i>		
						·		
				66.	Inter	rnational System for Human Cytogenetic Nomenclature (ISCN) compatible string:		
				67.		cify number of distinct cytogenetic abnormalities		
						One (1)		
						Two (2)		
						Three (3)		
						Four or more (4 or more)		
				68.	Spe	cify abnormalities (check all that apply)		
						-5		
						-7		
						-17		
						-18		
						-X		
						-Y		
						+4		
						+8		
						+11		
						+13		
						+14		
						+21		
						+22		
						t(3;3)		
						t(6;9)		

CIBMTR Center Nu	ımber:		CIBMTR Research ID:	
			t(9;11)	
			t(9;22)	
			t(15;17) and variants	
			t(16;16)	
			del(3q) / 3q-	
			del(5q) / 5q-	
			del(7q) / 7q-	
			del(9q) / 9q-	
			del(11q) / 11q-	
			del(16q) / 16q-	
			del(17q) / 17q-	
			del(20q) / 20q-	
			del(21q) / 21q-	
			inv(3)	
			inv(16)	
			(11q23) any abnormality	
			12p any abnormality	
			Other abnormality – <i>Go to question</i> 69	
		69.	Specify other abnormality:	
70.	Were cytoge	netics	s tested via karyotyping?	
☐ Yes – Go to q ☐ No – Go to q ☐ 71. Results of t			question 71	
			estion 76	
			ests	
	□ Abı	norma	alities identified – <i>Go to question 72</i>	
☐ No evalua			able metaphases – Go to question 76	
	□ No	abno	rmalities – Go to question 76	
	72.	Inter	rnational System for Human Cytogenetic Nomenclature (ISCN) compatible string:	
	73.	Spe	cify number of distinct cytogenetic abnormalities	
			One (1)	
			Two (2)	
			Three (3)	
			Four or more (4 or more)	

CIBMTR Ce	nter	Number: CIBMTR Research ID:
	76	Was documentation submitted to the CIBMTR? (e.g. cytogenetic or FISH report) ☐ Yes ☐ No
77.	Wei	e tests for molecular markers performed? (e.g. PCR, NGS) (at last evaluation)
		Yes – Go to question 78
		No – Go to question 90
		Unknown – Go to question 90
	78	CEBPA
		☐ Positive – Go to question 79
		☐ Negative – Go to question 80
		□ Not done – Go to question 80
		79. Specify CEBPA mutation
		☐ Biallelic (double mutant)
		☐ Monoallelic (single mutant)
		□ Unknown
	80	FLT3– TKD (point mutations in D835 or deletions of codon I836)
		□ Positive
		□ Negative
		□ Not done
	81	FLT3 – ITD mutation
		☐ Positive – Go to question 82
		☐ Negative – Go to question 84
		□ Not done – Go to question 84
		82. FLT3 – ITD allelic ratio
		☐ Known – Go to question 83
		☐ Unknown – Go to question 84
		83. Specify FLT3 - ITD allelic ratio:
	84	IDH1
		□ Positive
		□ Negative
		□ Not done

CIBMTR Center Number:		lumber: CIBMTR Research ID:			
	85.	IDH2			
		□ Positive			
		□ Negative			
		□ Not done			
	86.	KIT			
		□ Positive			
		□ Negative			
		□ Not done			
	87.	NPM1			
		□ Positive			
		□ Negative			
		□ Not done			
	Cop	by and complete questions 88-89 to report multiple other molecular markers			
	88.	Other molecular marker			
		☐ Positive – Go to question 89			
		☐ Negative – Go to question 89			
		□ Not done – Go to question 90			
		89. Specify other molecular marker:			
	Copy and complete questions 88-89 to report multiple other molecular markers				
CNS L	_euke	emia			
90.	Did the recipient have central nervous system leukemia at any time prior to the start of the preparative regimen / infusion?				
	□ \	/es			
	□ 1	No			
	п (Jnknown			
Status	at tı	ransplantation / infusion			
91.	What	t was the disease status? (based on hematological test results)			
	□ F	Primary induction failure – Go to question 103			
		st complete remission (no previous bone marrow or extramedullary relapse) (include CRi) – Go to question 92			
OIDMED E		2nd complete remission (include CRi) – Go to question 93			

CIBMTR Center	· Numbe	r: CIBMTR Research ID:					
	≥ 3rd c	complete remission (include CRi) – Go to question 93					
	1st rela	apse – Go to question 102					
	2nd rel	d relapse – Go to question 102					
	≥ 3rd r	elapse – Go to question 102					
	No trea	atment – Go to question 103					
92	2. Hov	w many cycles of induction therapy were required to achieve 1st complete remission? <i>(includes i)</i>					
		1					
		2					
		≥ 3					
93	3. Spe <i>app</i>	ecify method(s) that was used to assess measurable residual disease status (check all that oly)					
		FISH – Go to question 94					
		Karyotyping – Go to question 95					
		Flow cytometry – Go to question 96					
		PCR – Go to question 100					
		NGS – Go to question 101					
		Not assessed – Go to question 103					
	94.	Was measurable residual disease detected by FISH?					
		□ Yes					
		□ No					
	95.	Was measurable residual disease detected by karyotyping assay?					
		□ Yes					
		□ No					
	96.	Which leukemia immunophenotype was used for measurable residual disease detection? (check all that apply)					
		☐ Original leukemia immunophenotype – <i>Go to question</i> 97					
		☐ Aberrant phenotype – Go to question 98					
		97. Lower limit of detection (for the original leukemia immunophenotype):					
		98. Lower limit of detection (for the aberrant phenotype):					
	99.	Was measurable residual disease detected by flow cytometry?					
		□ Yes					

CIBMTR Cent	er N	umber: CIBMTR Research ID:
		□ No
		100. Was measurable residual disease detected by PCR? ☐ Yes ☐ No
		101. Was measurable residual disease detected by NGS? ☐ Yes ☐ No
	102	Date of most recent relapse:
103. D	ate	assessed:
Acute Lymph	obl	estic Leukemia (ALL)
104. S	Spec	fy ALL classification
B	_	nphoblastic leukemia / lymphoma -lymphoblastic leukemia / lymphoma, NOS (191)
]	-lymphoblastic leukemia / lymphoma with high hyperdiploidy (82)
]	-lymphoblastic leukemia / lymphoma with hypodiploidy (83)
]	-lymphoblastic leukemia / lymphoma, with <i>iAMP21</i> (95)
]	-lymphoblastic leukemia / lymphoma with <i>BCR::ABL1</i> fusion (192)
		-lymphoblastic leukemia / lymphoma, BCR::ABL1-like features (94)
		-lymphoblastic leukemia / lymphoma with <i>KMT2A</i> rearrangement (193)
		-lymphoblastic leukemia / lymphoma with <i>ETV6::RUNX1</i> fusion (195)
		-lymphoblastic leukemia / lymphoma with ETV6::RUNX1-like features (1111)
		-lymphoblastic leukemia / lymphoma with <i>TCF3::PBX1</i> fusion (194)
		-lymphoblastic leukemia / lymphoma with <i>IGH::IL3</i> fusion (81)
	J	-lymphoblastic leukemia / lymphoma with <i>TCF3::HLF</i> fusion (1112)
B	_	nphoblastic leukemia / lymphoma with other defined genetic abnormalities -lymphoblastic leukemia / lymphoma with DUX4 rearrangement (1113)
]	-lymphoblastic leukemia / lymphoma with <i>IG::MYC</i> fusion (1114)
]	-lymphoblastic leukemia / lymphoma with <i>MEF2D</i> rearrangement (1115)
]	-lymphoblastic leukemia / lymphoma with <i>ZNF384</i> rearrangement (1116)
]	-lymphoblastic leukemia / lymphoma with <i>NUTM1</i> rearrangement (1117)

CIBMTR Ce	nter	Number: CIBMTR Research ID:
		B-lymphoblastic leukemia / lymphoma with <i>PAX5alt</i> abnormalities (1118)
		B-lymphoblastic leukemia / lymphoma with PAX5 p.P80R abnormalities (1119)
	T-c	cell lymphoblastic leukemia / lymphoma T-lymphoblastic leukemia / lymphoma (196)
		Early T-precursor lymphoblastic leukemia / lymphoma (96)
		Early T-precursor lymphoblastic leukemia / lymphoma, with <i>BCL11B</i> rearrangement (1120)
	NK	Cell lymphoblastic leukemia / lymphoma Natural killer (NK)- cell lymphoblastic leukemia / lymphoma (97)
105.	Did	I the recipient have a predisposing condition?
		Yes – Go to question 106
		No – Go to question 108
		Unknown – Go to question 108
	10	06. Specify condition
		☐ Aplastic anemia – Also complete CIBMTR Form 2028 — APL – Go to question 108
		☐ Bloom syndrome – Go to question 108
		☐ Down syndrome – <i>Go to question 108</i>
		☐ Fanconi anemia – Also complete CIBMTR Form 2029 — FAN – Go to question 108
		☐ Other condition – Go to question 107
		107. Specify other condition:
108.		ere tyrosine kinase inhibitors given for therapy at any time prior to the start of the preparative regimen / fusion? (e.g. imatinib mesylate, dasatinib, etc.)
		Yes
		No
Laboi	ato	ry studies at diagnosis
109.	We	ere cytogenetics tested (karyotyping or FISH)? (at diagnosis)
		Yes – Go to question 110
		No – Go to question 123
		Unknown – Go to question 123
	11	10. Were cytogenetics tested via FISH?
		☐ Yes – Go to question 111
		□ No – Go to question 116

CIBMTR Center Number:			CIBMTR Research ID:
111.	Results of tests		
	☐ Abnormalities identified – <i>Go to question 112</i>		
	□ No abnormalities – <i>Go to question 116</i>		
	112.	Inter	rnational System for Human Cytogenetic Nomenclature (ISCN) compatible string
	113.	Spe	cify number of distinct cytogenetic abnormalities
			One (1)
			Two (2)
			Three (3)
			Four or more (4 or more)
	114.	Spe	cify abnormalities (check all that apply)
			-7
			+4
			+8
			+17
			+21
			t(1;19)
			t(2;8)
			t(4;11)
			t(5;14)
			t(8;14)
			t(8;22)
			t(9;22)
			t(10;14)
			t(11;14)
			t(12;21)
			del(6q) / 6q-
			del(9p) / 9p-
			del(12p) / 12p-
			add(14q)
			(11q23) any abnormality
			9p any abnormality
			12p any abnormality
			Hyperdiploid (> 50)

☐ Hypodiploid (< 46)

CIBMTR Center Number:	CIBMTR Research ID:
	□ iAMP21
	☐ Other abnormality – <i>Go to question 115</i>
	115. Specify other abnormality:
116. Were cytoge	enetics tested via karyotyping?
□ Yes – Go	to question 117
□ No – Go	to question 122
117. Result	ts of tests
□ Ab	normalities identified – <i>Go to question 118</i>
□ No	evaluable metaphases – Go to question 122
□ No	abnormalities – Go to question 122
118.	International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
119.	Specify number of distinct cytogenetic abnormalities
	□ One (1)
	□ Two (2)
	□ Three (3)
	☐ Four or more (4 or more)
120.	Specify abnormalities (check all that apply)
	□ -7
	□ +4
	□ +8
	□ +17
	□ + 21
	□ t(1;19)
	□ t(2;8)
	□ t(4;11)
	□ t(5;14)
	□ t(8;14)
	□ t(8;22)
	□ t(9;22)
	□ t(10;14)
	□ t(11;14)
	□ t(12;21)

CIBMTR Center Number:	CIBMTR Research ID:
	del(6q) / 6q-
	del(9p) / 9p-
	del(12p) / 12p-
	add(14q)
	(11q23) any abnormality
	9p any abnormality
	12p any abnormality
	Hyperdiploid (> 50)
	Hypodiploid (< 46)
	iAMP21
	Other abnormality – Go to question 121
121.	Specify other abnormality:
122. Was documentation	on submitted to the CIBMTR? (e.g. cytogenetic or FISH report)
☐ Yes	
□ No	
	markers performed? (e.g. PCR, NGS) (at diagnosis)
☐ Yes – Go to question	
□ No – Go to question	
□ Unknown – Go to qu	estion 120
124. BCR / ABL	
☐ Positive	
☐ Negative	
☐ Not done	
125. TEL-AML / AML1	
☐ Positive	
☐ Negative	
☐ Not done	
Copy and complete que	stions 126-127 for additional molecular markers
126. Other molecular m	narker
☐ Positive – Go t	to question 127
☐ Negative – <i>Go</i>	to question 127
□ Not done – Go	to question 128

CIBMTR Center	Number:		CIBMTR Research ID:
	127. Specif	fy othe	er molecular marker:
Со	py and complete	e ques	stions 126-127 for additional molecular markers
	ween diagnosis valuation for thi		ast evaluation (If subsequent infusion, labs after prior infusion and before sion)
128. We	re cytogenetics to	ested	(karyotyping or FISH)? (in between)
	Yes – Go to qu	ıestio	n 129
	No – Go to que	estion	142
	Unknown – <i>Go</i>	to qu	estion 142
12	29. Were cytoge	netics	tested via FISH?
	□ Yes – Go	to qu	uestion 130
	□ No – Go	to qu	estion 135
	130. Result	ts of te	ests
	□ Abı	norma	lities identified – Go to question 131
	□ No	abnor	rmalities – Go to question 135
	131.	Inter	national System for Human Cytogenetic Nomenclature (ISCN) compatible string:
	132.	Spec	cify number of distinct cytogenetic abnormalities
			One (1)
			Two (2)
			Three (3)
			Four or more (4 or more)
	133.	Spec	cify abnormalities <i>(check all that apply)</i>
			-7
			+4
			+8
			+17
			+21
			t(1;19)
			t(2;8)
			t(4;11)
			t(5;14)
			t(8;14)

CIBMTR Center Number:	CIBMTR Research ID:
Е	I t(8;22)
	l t(9;22)
	I t(10;14)
	l t(11;14)
	l t(12;21)
	l del(6q) / 6q-
С	l del(9p) / 9p-
	l del(12p) / 12p-
	l add(14q)
	l (11q23) any abnormality
	l 9p any abnormality
	l 12p any abnormality
	Hyperdiploid (> 50)
	l Hypodiploid (< 46)
	I iAMP21
С	Other abnormality – <i>Go to question 134</i>
13	4. Specify other abnormality:
135. Were cytogeneti	cs tested via karyotyping?
□ Yes – Go to	question 136
□ No – Go to c	uestion 141
136. Results of	tests
☐ Abnorr	nalities identified – <i>Go to question 137</i>
☐ No eva	luable metaphases – <i>Go to question 141</i>
☐ No abr	normalities – Go to question 141
137. Int	ernational System for Human Cytogenetic Nomenclature (ISCN) compatible string:
138. Sp	ecify number of distinct cytogenetic abnormalities
	l One (1)
	1 Two (2)
	Three (3)
	l Four or more (4 or more)
139. Sp	ecify abnormalities (check all that apply)
С	1 –7

CIBMTR Center Number:	CIBMTR Research ID:
	+4
	+8
	+17
	+21
	t(1;19)
	t(2;8)
	t(4;11)
	t(5;14)
	t(8;14)
	t(8;22)
	t(9;22)
	t(10;14)
	t(11;14)
	t(12;21)
	del(6q) / 6q-
	del(9p) / 9p-
	del(12p) / 12p-
	add(14q)
	(11q23) any abnormality
	9p any abnormality
	12p any abnormality
	Hyperdiploid (> 50)
	Hypodiploid (< 46)
	iAMP21
	Other abnormality – Go to question 140
140.	Specify other abnormality:
141. Was documentation	on submitted to the CIBMTR? (e.g. cytogenetic or FISH report)
☐ Yes	
□ No	
	markers performed? (e.g. PCR, NGS) (in between)
☐ Yes – Go to question	
□ No – Go to question	
□ Unknown – Go to qu	esuon 147

CIBMTR Center Nu	mber: CIBMTR Research ID:
143.	BCR / ABL
	□ Positive
	□ Negative
	□ Not done
144.	TEL-AML / AML1
	□ Positive
	□ Negative
	□ Not done
Сору	and complete questions 145-146 for additional molecular markers
145.	Other molecular marker
	□ Positive – Go to question 146
	□ Negative – Go to question 146
	□ Not done – Go to question 147
	146. Specify other molecular marker:
Сору	and complete questions 145-146 for additional molecular markers
Labora	ntory studies at last evaluation
147. Were o	ytogenetics tested (karyotyping or FISH)? (at last evaluation)
□ Ye	s – Go to question 148
□ No	- Go to question 161
□ Ur	known – Go to question 161
148.	Were cytogenetics tested via FISH?
	□ Yes – Go to question 149
	□ No – Go to question 154
	149. Results of tests
	☐ Abnormalities identified – <i>Go to question 150</i>
	☐ No abnormalities – <i>Go to question 154</i>
	150. International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
	151. Specify number of distinct cytogenetic abnormalities
	□ One (1)

CIBMTR Center Number:	CIBMTR Research ID:
	Two (2)
	Three (3)
	Four or more (4 or more)
·	cify abnormalities (check all that apply)
	-7
	+4
	+8
	+17
	+21
	t(1;19)
	t(2;8)
	t(4;11)
	t(5;14)
	t(8;14)
	t(8;22)
	t(9;22)
	t(10;14)
	t(11;14)
	t(12;21)
	del(6q) / 6q-
	del(9p) / 9p-
	del(12p) / 12p-
	add(14q)
	(11q23) any abnormality
	9p any abnormality
	12p any abnormality
	Hyperdiploid (> 50)
	Hypodiploid (< 46)
	iAMP21
	Other abnormality – <i>Go to question 153</i>
153.	Specify other abnormality:
154. Were cytogenetics	s tested via karyotyping?
□ Yes – Go to q	uestion 155

□ No – Go to question 160

CIBMTR Center Number:			CIBMTR Research ID:
155.	Result	s of to	ests
	☐ Abnormalities identified – <i>Go to question 156</i>		
	□ No	evalu	able metaphases – Go to question 160
	□ No	abno	rmalities – Go to question 160
	156.	Inte	national System for Human Cytogenetic Nomenclature (ISCN) compatible string:
	157.	Spe	cify number of distinct cytogenetic abnormalities
		_	One (1)
			Two (2)
			Three (3)
			Four or more (4 or more)
	158.	Spe	cify abnormalities (check all that apply)
			-7
			+4
			+8
			+17
			+21
			t(1;19)
			t(2;8)
			t(4;11)
			t(5;14)
			t(8;14)
			t(8;22)
			t(9;22)
			t(10;14)
			t(11;14)
			t(12;21)
			del(6q) / 6q-
			del(9p) / 9p-
			del(12p) / 12p-
			add(14q)
			(11q23) any abnormality
			9p any abnormality
			12p any abnormality

☐ Hyperdiploid (> 50)

CIBMTR Ce	nter Number: CIBMTR Research ID:
	☐ Hypodiploid (< 46)
	□ iAMP21
	☐ Other abnormality – <i>Go to question 159</i>
	159. Specify other abnormality:
	160. Was documentation submitted to the CIBMTR? (e.g. cytogenetic or FISH report)
	□ Yes
	□ No
161.	Were tests for molecular markers performed? (e.g. PCR, NGS) (at last evaluation)
	☐ Yes – Go to question 162
	□ No – Go to question 166
	□ Unknown – Go to question 166
	162. BCR / ABL
	□ Positive
	□ Negative
	□ Not done
	163. TEL-AML / AML1
	□ Positive
	□ Negative
	□ Not done
	Copy and complete questions 164-165 for additional molecular markers
	164. Other molecular marker
	☐ Positive – Go to question 165
	☐ Negative – Go to question 165
	□ Not done – Go to question 166
	165. Specify other molecular marker:
	Copy and complete questions 164-165 for additional molecular markers
	CNS Leukemia
166.	Did the recipient have central nervous system leukemia at any time prior to the start of the preparative regimen / infusion?
	□ Yes

CIBMTR	Cen	ter I	Number: CIBMTR Research ID:
	[No
	[Unknown
	•	Stat	us at transplantation / infusion
16	67. \	Nha	t was the disease status? (based on hematological test results)
	[Primary induction failure – Go to question 179
	[1st complete remission (no previous bone marrow or extramedullary relapse) (include CRi) – Go to question 168
	[2nd complete remission (include CRi) – Go to question 169
	[≥ 3rd complete remission (include CRi) – Go to question 169
	[1st relapse – <i>Go to question 178</i>
	[2nd relapse – <i>Go to question 178</i>
	[≥ 3rd relapse – <i>Go to question 178</i>
	[No treatment – <i>Go to question 179</i>
		168	How many cycles of induction therapy were required to achieve 1st complete remission? (includes CRi) — .
			□≥3
		169	 Specify method(s) that was used to assess measurable residual disease status (check all that apply)
			□ FISH – Go to question 170
			☐ Karyotyping – <i>Go to question 171</i>
			☐ Flow cytometry – Go to question 172
			□ PCR – Go to question 176
			□ NGS – Go to question 177
			□ Not assessed – Go to question 179
			170. Was measurable residual disease detected by FISH?
			□ Yes
			□ No
			171. Was measurable residual disease detected by karyotyping assay? ☐ Yes
			□ No

CIBMTR Center N	Number:	: CIBMTR Research ID:							
	172.	 Which leukemia immunophenotype was used for measurable residual disease detection? (check all that apply) □ Original leukemia immunophenotype – Go to question 173 							
		□ Abe	errant phenoty	ype – Go to qu	estion 1	74			
		173. Lower limit of detection (for the original leukemia immunophenotype):							
		174.	Lower limit o	of detection (for	the aberra	ant phenot	type):		
	175.	Was m	neasurable res	sidual disease o	letected b	by flow cyto	ometry?		
		☐ Yes	;						
		□ No							
	176.	Was m	neasurable res	sidual disease d	letected k	y PCR?			
		☐ Yes	3						
		□ No							
	177.	Was m	neasurable res	sidual disease d	detected b	y NGS?			
		☐ Yes	3						
		□ No							
179	R Date	of most	recent relaps	· • ·			_		
170	Date (Ji iiiost	recent relaps	YYYY			DD		
470 D-4-		. al .				0- 4			
179. Date	assesse	ea:	YYYY	. — <u> </u>		- Go to er	nd of form		
Acute Leukemia	s of Mix	ed or A	mbiquous Li	ineage					
				9					
180. Spec	cify acute	leuker	nias of mixed	or ambiguous I	ineage				
	Blastic pl	asmacy	ytoid dendritic	cell neoplasm	(296) – G	o to ques	stion 182		
	Acute un	differen	itiated leukem	nia (31) – Go to	questio	n 182			
	Mixed ph	enotyp	e acute leuke	mia (MPAL) wit	h BCR::A	BL1 fusior	n (84) – Go to d	question 182	
	Mixed ph	enotyp	e acute leuke	mia with KMT2	۲ rearranç	gement (8	5) – Go to que	stion 182	
	Mixed-phenotype acute leukemia with ZNF384 rearrangement (1051) – <i>Go to question 182</i>						juestion 182		
	Acute leukemia of ambiguous lineage with BCL11B rearrangement (1052) – Go to question					to question 182			
	Mixed-ph	enotyp	e acute leuke	mia, B / myeloid	1 (86) – G	o to que:	stion 182		
	Mixed-ph	enotyp	e acute leuke	mia, T / myeloid	i (87) – G	io to que:	stion 182		
	Mixed-ph	enotyp	e acute leuke	mia, rare types	(1053) –	Go to que	estion 182		
	Acute leu	ıkemia	of ambiguous	lineage, NOS ((88) – Go	to quest	ion 181		

CIBMTR Center	Number: CIBMTR Research ID:
18	Specify other acute leukemia of ambiguous lineage or myeloid neoplasm:
Stat	us at transplantation / infusion
182. Wha	t was the disease status? (based on hematological test results)
	Primary induction failure
	1st complete remission (no previous marrow or extramedullary relapse)
	2nd complete remission
	≥ 3rd complete remission
	1st relapse
	2nd relapse
	≥3rd relapse
	No treatment
183	3. Date assessed:
	YYYY MM DD
Chronic Myeloid	I Leukemia (CML)
184. Was	therapy given prior to this HCT?
	Yes – Go to question 185
	No – Go to question 191
18	5. Combination chemotherapy
	□ Yes
	□ No
180	6. Hydroxyurea (Droxia, Hydrea)
	□ Yes
	□ No
18	7. Tyrosine kinase inhibitor (e.g. imatinib mesylate, dasatinib, nilotinib)
	□ Yes
	□ No
18	3. Interferon-α (Intron, Roferon) (includes PEG)
	□ Yes
	□ No
189	9. Other therapy

CIBMTR Ce	nter	Number: CIBMTR Research ID:
		☐ Yes – Go to question 190
		□ No – Go to question 191
		190. Specify other therapy:
191.	Wh	nat was the disease status?
		Complete hematologic response (CHR) preceded only by chronic phase – Go to question 192
		Complete hematologic response (CHR) preceded by accelerated phase and/or blast phase – Go to question 192
		Chronic phase – Go to question 192
		Accelerated phase – Go to question 193
		Blast phase – Go to question 193
	19	92. Specify level of response
		☐ No cytogenetic response (No CyR)
		☐ Minimal cytogenetic response
		☐ Minor cytogenetic response
		☐ Partial cytogenetic response (PCyR)
		☐ Complete cytogenetic response (CCyR)
		☐ Major molecular remission (MMR)
		☐ Complete molecular remission (CMR)
400	N 1	
193.		mber
		1st
		2nd
		3rd or higher
194.	Dat	te assessed:
		YYYY MM DD
Myelodyspl	asti	ic Syndrome (MDS)
195.		nat was the MDS subtype at diagnosis? – If transformed to AML, indicate AML as primary disease; so complete AML Disease Classification questions
	MD	OS with defining genetic abnormalities Myelodysplastic syndrome with low blasts and isolated 5q deletion (MDS-5q) (66) – Go to question 198
		Myelodysplastic syndrome with low blasts and <i>SF3B1</i> mutation (MDS-SF3B1) (1411) – <i>Go to</i> question 198

CIBMTR Ce	nter	Number: CIBMTR Research ID:
		Myelodysplastic syndrome with low blasts and ring sideroblasts (>=15% ring sideroblasts and wild type SF3B1) (1412) – <i>Go to question 198</i>
		Myelodysplastic syndrome with biallelic <i>TP53</i> inactivation (MDS-biTP53) (1413) – <i>Go to question</i> 198
	MD	S, morphically defined MDS, with low blasts (MDS-LB; <5% BM, <2%PB) (1414) – Go to question 198
		MDS, hypoplastic (MDS-h) <=25% cellularity by age (1415) – <i>Go to question 198</i>
		MDS with increased blasts (MDS-IB1) (61) - Go to question 198
		MDS with increased blasts (MDS-IB2) (62) – Go to question 198
		MDS with fibrosis (MDS-f) (1416) – Go to question 198
	Chi	ildhood myelodysplastic neoplasms (MDS) Childhood MDS with low blasts, hypocellular (68) – Go to question 198
		Childhood MDS with low blasts, not otherwise specified (1417) – Go to question 198
		Childhood MDS with increased blasts (1418) – <i>Go to question 198</i>
	My	elodysplastic / myeloproliferative neoplasms Chronic myelomonocytic leukemia (CMML), Myelodysplastic (54) – Go to question 198
		Chronic myelomonocytic leukemia (CMML), Myeloproliferative (1419) – Go to question 198
		Myelodysplastic/myeloproliferative neoplasm with SF3B1 mutation and thrombocytosis (1452) – $\textbf{\textit{Go to}}$ $\textbf{\textit{question 198}}$
		MDS/MPN with ring sideroblasts (>=15% ring sideroblasts and wild type SF3B1) and thrombocytosis (1420) – <i>Go to question 198</i>
		Juvenile myelomonocytic leukemia (JMML) (36) – Go to question 234
		Myelodysplastic/myeloproliferative neoplasm with neutrophilia – (1440) – <i>Go to question</i> 392
		Myelodysplastic syndrome / myeloproliferative neoplasm, NOS (69) – <i>Go to question 197</i>
	19	6. Specify Myelodysplastic syndrome, unclassifiable (MDS-U)
		☐ MDS-U with 1% blood blasts
		☐ MDS-U with single lineage dysplasia and pancytopenia
		☐ MDS-U based on defining cytogenetic abnormality
	19	7. Was documentation submitted to the CIBMTR? (e.g. pathology report used for diagnosis)
		□ Yes
		□ No
198.	Wa	s the disease MDS therapy related?
		Yes
		No
		Unknown

CIBMTR Cente	er Number:	CIBMTR Research ID:
199. Di	d the recipient have a predisposing	g condition?
	Yes – Go to question 200	
	No - Go to question 202	
	Unknown – Go to question 20	2
2	200. Specify condition	
	☐ Aplastic anemia – Also co	omplete CIBMTR Form 2028 – APL – Go to question 202
	☐ DDX41-associated familia	ll MDS – Go to question 202
	☐ Diamond-Blackfan Anemia	a – Go to question 202
	☐ Fanconi anemia – Go to o	question 202
	☐ GATA2 deficiency (includi to question 202	ing Emberger syndrome, MonoMac syndrome, DCML deficiency) – Go
	☐ Li-Fraumeni Syndrome – 0	Go to question 202
	☐ Paroxysmal nocturnal hen question 202	noglobinuria – Also complete CIBMTR Form 2028 – APL – Go to
	☐ RUNX1 deficiency (previo malignancies") – Go to qu	usly "familial platelet disorder with propensity to myeloid uestion 202
	☐ SAMD9- or SAMD9L-asso	ociated familial MDS – <i>Go to question 202</i>
	☐ Shwachman-Diamond Syr	ndrome – Go to question 202
	☐ Telomere biology disorder 2028 – APL – Go to que:	r (including dyskeratosis congenita) – Also complete CIBMTR Form stion 202
	☐ Other condition – Go to q	uestion 201
	201. Specify other condition	1:
Laborate	ory studies at diagnosis of MDS	
202. Da	ate CBC drawn:	
203. W	ВС	
	Known – Go to question 204	
	Unknown – Go to question 20	5
2	204•_	
		$\Box \times 10^9$ /L (x 10^3 /mm ³)
		□ x 10 ⁶ /L
205. Ne	eutrophils	
	Known – Go to question 206	
	Unknown – Go to question 20	7

CIBMTR Center Number:	CIBMTR Research ID:
206%	
207. Blasts in blood	
☐ Known – Go to question 208	
☐ Unknown— Go to question 209	
208 %	
209. Hemoglobin	
☐ Known – Go to question 210	
☐ Unknown – Go to question 212	
210 •	
	□ g/dL
	□ g/L
	□ mmol/L
211. Were RBCs transfused ≤ 30 da	ays before date of test?
☐ Yes	
□ No	
212. Platelets	
☐ Known – Go to question 213	
☐ Unknown – Go to question 215	
213.	
	$\Box \times 10^9$ /L (x 10^3 /mm ³)
	□ x 10 ⁶ /L
214. Were platelets transfused ≤ 7 of	days before date of test?
☐ Yes	
□ No	
215. Blasts in bone marrow	
☐ Known – Go to question 216	
☐ Unknown – Go to question 217	
216 %	

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217. Were cytogenetics tested (karyotyping or FISH)?

CIBMTR Center Number:	CIBMTR Research ID:			
☐ Yes – Go to qu	uestion 218			
□ No – Go to que	estion 234			
□ Unknown – <i>Go</i>	known – Go to question 234			
040 Ware extens	metics tested via FICUS			
	enetics tested via FISH? o to question 219			
	to question 219			
□ No – Go	to question 220			
219. Samp	le source			
□ Blo	ood			
□ Во	ne marrow			
220. Resul	ts of tests			
	normalities identified – <i>Go to question 221</i>			
	abnormalities – <i>Go to question 225</i>			
Specif	y cytogenetic abnormalities identified via FISH at diagnosis			
221.	International System for Human Cytogenetic Nomenclature (ISCN) compatible string:			
222.	Specify number of distinct cytogenetic abnormalities			
	□ One (1)			
	□ Two (2)			
	□ Three (3)			
	☐ Four or more (4 or more)			
223.	Specify abnormalities (check all that apply)			
Mo	onosomy			
	□ -5 			
	□ -7			
	□ −13 □ −20			
	□ -20 □ -Y			
	□ -1			
Tri	somy			
	□ +8 □ 40			
	□ +19			

CIBMTR Center Number:	CIBMTR Research ID:
	t(1;3)
	t(2;11)
	t(3;3)
	t(3;21)
	t(6;9)
	t(11;16)
Deletion	
_	del(3q) / 3q-
	del(5q) / 5q-
	del(7q) / 7q-
	del(9q) / 9q-
	del(11q) / 11q-
	del(12p) / 12p-
	del(13q) / 13q-
	del(20q) / 20q-
Inversio	n
	inv(3)
Other	
	i17q
	Other abnormality – <i>Go to question 224</i>
224.	Specify other abnormality:
225. Was docum	entation submitted to the CIBMTR? (e.g. FISH report)
☐ Yes	
□ No	
226. Were cytogenetics	s tested via karyotyping?
☐ Yes – Go to q	uestion 227
□ No – Go to qu	restion 234
227. Sample sou	rce
☐ Blood	
☐ Bone ma	arrow
228. Results of to	ests
☐ Abnorma	alities identified – Go to question 229

CIBMTR Center Number:			CIBMTR Research ID:	
	□ No	evalu	uable metaphases – Go to question 233	
	□ No abnormalities – <i>Go to question 233</i>			
	Specify	/ cyto	ogenetic abnormalities identified via conventional cytogenetics at diagnosis	
	229.	Inte	national System for Human Cytogenetic Nomenclature (ISCN) compatible string:	
	230.	Spe	cify number of distinct cytogenetic abnormalities	
			One (1)	
			Two (2)	
			Three (3)	
			Four or more (4 or more)	
	231.	Spe	cify abnormalities (check all that apply)	
	Мо	noso	omy _5	
			-3 -7	
			–20	
			-Y	
	Tri	somy		
		_	+8	
			+19	
	Tra	nslo	cation	
			t(1;3)	
			t(2;11)	
			t(3;3)	
			t(3;21)	
			t(6;9)	
			t(11;16)	
	De	letion		
			del(3q) / 3q-	
			del(5q) / 5q-	
			del(7q) / 7q- del(9q) / 9q-	
			del(9q) / 9q- del(11q) / 11q-	
			α σι (ε τη <i>) /</i>	

CIBMTR Center Number:	CIBMTR Research ID:
	del(12p) / 12p-
	del(13q) / 13q-
	del(20q) / 20q-
Inversion	nv(3)
	111(3)
Other	147
	i17q
	Other abnormality – Go to question 232
232.	Specify other abnormality:
233. Was documentatio	n submitted to the CIBMTR? (e.g. karyotyping report)
□ Yes	
□ No	
234. Did the recipient progress of the preparative regime	or transform to a different MDS subtype or AML between diagnosis and the start n / infusion?
☐ Yes – Go to question	235
□ No – Go to question	239
235. Specify the MDS so	ubtype or AML after transformation
MDS with defining genet ☐ Myelodysplastic question 237	ic abnormalities syndrome with low blasts and isolated 5q deletion (MDS-5q) (66) – Go to
☐ Myelodysplastic question 237	syndrome with low blasts and <i>SF3B1</i> mutation (MDS-SF3B1) (1411) – <i>Go to</i>
	syndrome with low blasts and ring sideroblasts (>=15% ring sideroblasts and I) (1412) – <i>Go to question 237</i>
☐ Myelodysplastic question 237	syndrome with biallelic <i>TP53</i> inactivation (MDS-biTP53) (1413) – Go to
MDS, morphically defined □ MDS, with low	d blasts (MDS-LB; <5% BM, <2%PB) (1414) – <i>Go to question 237</i>
☐ MDS, hypoplast	tic (MDS-h) <=25% cellularity by age (1415) – <i>Go to question</i> 237
☐ MDS with increa	ased blasts (MDS-IB1) (61) – Go to question 237
☐ MDS with increa	ased blasts (MDS-IB2) (62) – Go to question 237
☐ MDS with fibros	is (MDS-f) (1416) – Go to question 237
Childhood myelodysplas ☐ Childhood MDS	vitic neoplasms (MDS) with low blasts, hypocellular (68) – <i>Go to question 237</i>

CIBMTR Center Number:	CIBMTR Resea	rch ID:						
☐ Childhood	☐ Childhood MDS with low blasts, not otherwise specified (1417) – <i>Go to question 237</i>							
☐ Childhood	MDS with increased blasts (1418	B) – Go to que	estion 237					
	veloproliferative neoplasms nyelomonocytic leukemia (CMML),	Myelodysplas	stic (54) – Go	to question	237			
☐ Chronic m	yelomonocytic leukemia (CMML),	Myeloprolifer	ative (1419) -	Go to ques	tion 237			
☐ Myelodysp	☐ Myelodysplastic/myeloproliferative neoplasm with neutrophilia (1440) – <i>Go to question</i> 237							
	☐ Myelodysplastic/myeloproliferative neoplasm with <i>SF3B1</i> mutation and thrombocytosis (1452) – <i>Go to question 237</i>							
	☐ MDS/MPN with ring sideroblasts (>=15% ring sideroblasts and wild type SF3B1) and thrombocytosis (1420) – <i>Go to question 237</i>							
☐ Myelodysp	olastic syndrome / myeloproliferati	ive neoplasm,	NOS (69) – (Go to questic	on 237			
Transformed to AM ☐ Transform	L aed to AML (70) – Go to question	238						
236. Specify	y Myelodysplastic syndrome, uncl	assifiable (MD	S-U)					
	S-U with 1% blood blasts – Go to	question 23	37					
	S-U with single lineage dysplasia	and pancytop	enia – Go to	question 23	7			
	S-U based on defining cytogenetic	c abnormality	– Go to que:	stion 237				
	y the date of the most recent trans to question 239	formation:						
			YYYY	MM	DD			
238 Date of	f MDS diagnosis:		– G	o to end of fo	rm			
200. Bate 6.	YYYY	MM	DD	0 (0 0)14 01 10				
•	t evaluation prior to the start of	the preparati	ve regimen /	infusion				
239. Date CBC drawn:		DD						
240. WBC								
☐ Known – Go to	question 241							
☐ Unknown – Go t	o question 242							
241	•							
		10 ³ /mm ³)						
	□ x 10 ⁶ /L							

242. Neutrophils

CIBMTR Center Number:	CIBMTR Research ID:
☐ Known – Go to question 243	
☐ Unknown – Go to question 24	4
243%	
244. Blasts in blood	
☐ Known – Go to question 245	
☐ Unknown – Go to question 24	6
245	
245 %	
246. Hemoglobin	
☐ Known – Go to question 247	
☐ Unknown – Go to question 24	9
247 • •	
	□ g/dL
	□ g/L
	□ mmol/L
248. Were RBCs transfused ≤ 30 o	days before date of test?
□ Yes	adyo pololo dalo ol toot.
□ No	
249. Platelets	
☐ Known – Go to question 250	2
☐ Unknown – Go to question 25.	2
250	_
	$\square \times 10^9 / L (x \cdot 10^3 / mm^3)$
	□ x 10 ⁶ /L
251. Were platelets transfused ≤ 7	days before date of test?
□ Yes	
□ No	
252. Blasts in bone marrow	
☐ Known – Go to question 253	
☐ Unknown – Go to question 25	4

CIBMTR Center Number:	CIBMTR Research ID:
253	%
254. Were cytogenetics to	ested (karyotyping or FISH)?
☐ Yes – Go to q	uestion 255
□ No – Go to qu	uestion 271
☐ Unknown – Go	to question 271
255. Were cytoge	netics tested via FISH?
☐ Yes – Go	to question 256
□ No – Go	to question 263
256. Sampl	le source
□ Blo	ood
□ Вог	ne marrow
257. Result	ts of tests
□ Abi	normalities identified – <i>Go to question 258</i>
□ No	abnormalities – Go to question 262
	y cytogenetic abnormalities identified via FISH at last evaluation prior to the start preparative regimen / infusion
258.	International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
259.	Specify number of distinct cytogenetic abnormalities
	□ One (1)
	□ Two (2)
	☐ Three (3)
	☐ Four or more (4 or more)
260.	Specify abnormalities (check all that apply)
Mo	onosomy
	□ -5
	□ -7
	□ -13
	□ –20
	□ -Y

CIBMTR Center Number: _		CIBMTR Research ID:
		+8
		+19
	Transloo	cation t(1;3)
		t(2;11)
		t(3;3)
		t(3;21)
		t(6;9)
		t(11;16)
	Deletion	del(3q) / 3q-
		del(5q) / 5q-
		del(7q) / 7q-
		del(9q) / 9q-
		del(11q) / 11q-
		del(12p) / 12p-
		del(13q) / 13q-
		del(20q) / 20q-
	Inversio	n inv(3)
	Other	
		i17q
		Other abnormality – <i>Go to question 261</i>
	261.	Specify other abnormality:
262. Was d	ocumentatio	on submitted to the CIBMTR? (e.g. FISH report)
☐ Yes	5	
□ No		
263. Were o	cytogenetics	tested via karyotyping?
☐ Yes	s – Go to qu	uestion 264
□ No	– Go to qu	estion 271
264.	Sample sou	rce
	□ Blood	

CIBMTR Center Number:		CIBMTR Research ID:	
265.	Results of	f tests	
☐ Abnormalities identified – <i>Go to question 266</i>			
	☐ No evaluable metaphases – <i>Go to question 270</i>		
☐ No abnormalities – <i>Go to question 270</i>			
		netic abnormalities identified via conventional cytogenetics at last evaluation art of the preparative regimen / infusion	
	266. In	ternational System for Human Cytogenetic Nomenclature (ISCN) compatible string:	
	267. Sp	pecify number of distinct cytogenetic abnormalities	
		One (1)	
		J Two (2)	
		Three (3)	
		Four or more (4 or more)	
	268. Sp	pecify abnormalities (check all that apply)	
	Monos	somy] _5	
] –13	
] –20	
] _Y	
	Trison		
		1 +8	
	L] +19	
		location 1 t(1;3)	
] t(2;11)	
		1 t(3;3)	
] t(3;21)	
		1 t(6;9)	
] t(11;16)	
	Deletio	on	
		del(3q) / 3q-	
		del(5q) / 5q-	

CIBMTR Center Number:	CIBMTR Research ID:
	del(7q) / 7q-
	del(9q) / 9q-
	del(11q) / 11q-
	del(12p) / 12p-
	del(13q) / 13q-
	del(20q) / 20q-
Inversion	n inv(3)
Other	
	i17q
	Other abnormality – Go to question 269
269.	Specify other abnormality:
270. Was documentatio ☐ Yes ☐ No	on submitted to the CIBMTR? (e.g. karyotyping report)
Status at transplantation	n / infusion
271. What was the disease sta	tus?
☐ Complete remission (CR) – Go to question 274
☐ Hematologic improver	ment (HI) – Go to question 272
☐ No response (NR) / st	table disease (SD) – Go to question 274
☐ Progression from hem	natologic improvement (Prog from HI) – <i>Go to question 274</i>
☐ Relapse from complete	te remission (Rel from CR) – Go to question 274
□ Not assessed – Go to	o end of form
272. Specify the cell line	es examined to determine HI status (check all that apply)
☐ HI-E – Go to q	uestion 273
☐ HI-P – Go to q	uestion 274
☐ HI-N – Go to q	uestion 274
273. Specify tran	sfusion dependence
☐ Non trans	sfused (NTD) – Go to question 274
☐ Low trans	sfusion burden (LTB) - Go to question 274
274. Date assessed:	

CIBMTR Cente	Number: CIBMTR Research ID:
Myeloprolifera	tive Neoplasms (MPN)
	nat was the MPN subtype at diagnosis? – If transformed to AML, indicate AML as primary diseas so complete AML Disease Classification questions
My	reloproliferative neoplasms Chronic neutrophilic leukemia (165) – <i>Go to question 278</i>
	Chronic eosinophilic leukemia (166) – <i>Go to question 278</i>
	Essential thrombocythemia (58) – <i>Go to question 278</i>
	Myeloproliferative neoplasm, NOS (60) – <i>Go to question 277</i>
	Polycythemia vera (PCV) (57) – <i>Go to question 278</i>
	Primary myelofibrosis (PMF) (167) – <i>Go to question</i> 278
Ma	estocytosis Cutaneous mastocytosis (CM) (1465) – <i>Go to question 278</i>
	Mast cell sarcoma (MCS) (1466) – Go to question 278
	Systemic mastocytosis (1470) – <i>Go to question 276</i>
2	76. Specify systemic mastocytosis
	☐ Indolent systemic mastocytosis (ISM) – <i>Go to question 278</i>
	☐ Smoldering systemic mastocytosis (SSM) – <i>Go to question 278</i>
	☐ Systemic mastocytosis with an associated hematological neoplasm (SM-AHN) – Go to question 278
	☐ Aggressive systemic mastocytosis (ASM) – <i>Go to question 278</i>
	☐ Mast cell leukemia (MCL) – Go to question 278
	☐ Bone marrow mastocytosis – <i>Go to question 278</i>
2	77. Was documentation submitted to the CIBMTR? (e.g. pathology report used for diagnosis) ☐ Yes ☐ No
Assessr	nent at diagnosis
	d the recipient have constitutional symptoms in six months before diagnosis? (symptoms are >10% eight loss in 6 months, night sweats, or unexplained fever higher than 37.5 °C)
	Yes
	No
	Unknown

Laboratory studies at diagnosis of MPN

CIBMTR Center Number:		CIBMTR Research ID:		
279.	Date CBC drawn:		 DD	
280.	WBC			
	☐ Known – Go to question 281			
	☐ Unknown – Go to question 282			
	·			
	281•	_		
			(x 10 ³ /mm ³)	
		□ x 10 ⁶ /L		
282.	Neutrophils			
	☐ Known – Go to question 283			
	☐ Unknown – Go to question 284			
	283%			
284.	Blasts in blood			
	☐ Known – Go to question 285			
	☐ Unknown— Go to question 286			
	285 %			
286.	Hemoglobin			
	☐ Known – Go to question 287			
	☐ Unknown – Go to question 289			
	287 •			
		□ g/dL		
		□ g/L		
		□ mmol/L		
	288. Were RBCs transfused ≤ 30 da	ıvs before dat	te of test?	
	□ Yes	,		
	□ No			
000	Districts			
∠89.	Platelets ☐ Known – Go to question 290			
	☐ Known - Go to question 290☐ Unknown - Go to question 292			
	D OIMIOWII – GO to question 292			

CIBMTR Center Nu	mber: CIBMTR Research ID:
	□ x 10 ⁹ /L (x 10 ³ /mm ³)
	□ x 10 ⁶ /L
291	Were platelets transfused ≤ 7 days before date of test?
	□ Yes
	□ No
292. Blasts	in bone marrow
	nown – Go to question 293
	sknown – Go to question 294
293.	%
294. Were to	ests for driver mutations performed?
□ Ye	s – Go to question 295
□ No	– Go to question 305
□ Un	known – Go to question 305
295.	JAK2
	□ Positive – Go to question 296
	□ Negative – Go to question 298
	□ Not done – Go to question 298
	296. JAK2 V617F
	□ Positive
	□ Negative
	□ Not done
	297. JAK2 Exon 12
	□ Positive
	□ Negative
	□ Not done
298.	CALR
	□ Positive – Go to question 299
	□ Negative – Go to question 302
	□ Not done – Go to question 302
	299. CALR type 1

CIBMTR Center Nur	mber: CIBMTR Research ID:
	□ Negative
	□ Not done
	200 CALB t 2
	300. CALR type 2 ☐ Positive
	☐ Negative
	☐ Not done
	301. Not defined
	□ Positive
	□ Negative
	☐ Not done
302.	MPL
	□ Positive
	□ Negative
	□ Not done
303.	CSF3R
	□ Positive
	□ Negative
	□ Not done
304.	Was documentation submitted to the CIBMTR?
	□ Yes
	□ No
305 Wara c	ytogenetics tested (karyotyping or FISH)?
	s – Go to question 306
	- Go to question 322
	known – Go to question 322
306.	Were cytogenetics tested via FISH?
	□ Yes – Go to question 307
	□ No – Go to question 314
	307. Sample source
	□ Blood
	☐ Bone marrow

CIBMTR Center Number:			CIBMTR Research ID:		
308.	Results	s of te	ests		
	☐ Abnormalities identified – <i>Go to question 309</i>				
	☐ No abnormalities – <i>Go to question 313</i>				
Speci	ify cyto	gene	tic abnormalities identified via FISH at diagnosis		
	309.	Inter	national System for Human Cytogenetic Nomenclature (ISCN) compatible string:		
	310.	Spec	cify number of distinct cytogenetic abnormalities		
			One (1)		
			Two (2)		
			Three (3)		
			Four or more (4 or more)		
	311.	Spe	cify abnormalities (check all that apply)		
	Mo	noso	my _5		
			-7		
			_Y		
	Tris	somy			
			+9		
	Tra	nslo	cation t(1;any)		
			t(3q21;any)		
			t(11q23;any)		
			t(12p11.2;any)		
			t(6;9)		
	Del	etion	del(5q) / 5q-		
			del(7q) / 7q-		
			del(11q) / 11q-		
			del(12p) / 12p-		
			del(13q) / 13q-		
			del(20q) / 20q-		

CIBMTR Center Number: _	CIBMTR Research ID:
	Inversion
	□ dup(1)
	□ inv(3)
	Other
	□ i17q
	☐ Other abnormality – Go to question 312
	312. Specify other abnormality:
313. V	Nas documentation submitted to the CIBMTR? (e.g. FISH report)
I	□ Yes
!	□ No
314. Were c	ytogenetics tested via karyotyping?
☐ Yes	– Go to question 315
□ No -	- Go to question 322
315. \$	Sample source
1	□ Blood
I	□ Bone marrow
316. F	Results of tests
1	☐ Abnormalities identified – <i>Go to question 317</i>
I	□ No evaluable metaphases – <i>Go to question 321</i>
1	□ No abnormalities – <i>Go to question 321</i>
Specify	y cytogenetic abnormalities identified via conventional cytogenetics at diagnosis
	317. International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
	318. Specify number of distinct cytogenetic abnormalities
	□ One (1)
	□ Two (2)
	☐ Three (3)
	☐ Four or more (4 or more)
	319. Specify abnormalities (check all that apply)
	Monosomy
	□ –5

CIBMTR Center Number:	CIBMTR Research ID:
	-7
	-Y
Trisomy	
	+9
Transloo	t(1;any)
	t(3q21;any)
_	t(11q23;any)
	t(12p11.2;any)
	t(6;9)
Deletion □	del(5q) / 5q-
	del(7q) / 7q-
_	del(11q) / 11q-
	del(12p) / 12p-
	del(13q) / 13q-
	del(20q) / 20q-
Inversio	n dup(1)
	inv(3)
Other	i17q
	Other abnormality – <i>Go to question 320</i>
320.	Specify other abnormality:
321. Was docum	entation submitted to the CIBMTR? (e.g. karyotyping report)
☐ Yes	
□ No	
322. Did the recipient progress of the preparative regime	or transform to a different MPN subtype or AML between diagnosis and the start in / infusion?
☐ Yes – Go to questio	n 323
□ No – Go to questior	326

CIBMTR Ce	enter	Number	:		CIBMTR Rese	earch ID:			
		ПΡ	ost-essential th	nrombocyth	emic myelofibr	osis (1467) – G	o to questio	on 324	
		ПΡ	ost-polycythen	nic myelofib	rosis (1468) –	Go to questio	n 324		
		ПΤ	ransformed to	AML (70) –	Go to questi	on 325			
		324.	. Specify the d - Go to que			nsformation:			– DD
		225	Data of MDN	diagnasis			0	o to end of fo	
		320.	. Date of MPN	diagnosis.			DD	o to ena or ro)
	As	sessmen	nt at last evalu	ation prior	to the start o	f the preparati	ve regimen/ i	nfusion	
326.	Sp	ecify trans	sfusion depend	lence at las	t evaluation pr	ior to the start o	of the preparat	tive regimen/ ir	nfusion
		Non-tra	nsfused (NTD)	- (0 RBCs	in 16 weeks)				
		Low-transfusion burden (LTB) – (3-7 RBCs in 16 weeks in at least 2 transfusion episodes; maximum of 3 in 8 weeks)							
		High-tra	insfusion burde	en (HTB) – (≥ 8 RBCs in 1	6 weeks; ≥ 4 ii	n 8 weeks)		
327.	pr	eparative		sion? <i>(symp</i>	otoms are >10	months before % weight loss in		-	
		Yes							
		No							
		Unknow	/n						
328.		I the recip	pient have sple	nomegaly a	ıt last evaluatio	on prior to the st	art of the prep	parative regime	en/
		Yes – G	o to question	329					
		No – G o	o to question	332					
		Unknow	n – Go to qu e	estion 332					
		Not app	licable (splene	ctomy) – G	o to question	332			
	32	29. Spec	cify the method	used to me	easure spleen :	size			
		ΠР	hysical assess	ment – Go	to question 3	330			
		□U	lltrasound – Go	to questi	on 331				
		□С	T/ MRI – Go t o	o question	331				
		330.	. Specify the s	pleen size:	centir	neters below le	ft costal marg	in – Go to qu e	estion 332
		331.	. Specify the s	pleen size:	centir	neters			

CIBMTR Ce	enter Number: CIBMTR Research ID:
332.	Did the recipient have hepatomegaly at last evaluation prior to the start of the preparative regimen / infusion?
	☐ Yes – Go to question 333
	□ No – Go to question 336
	□ Unknown – Go to question 336
	333. Specify the method used to measure liver size
	☐ Physical assessment – <i>Go to question 334</i>
	☐ Ultrasound – Go to question 335
	□ CT/ MRI – Go to question 335
	334. Specify the liver size: centimeters below right costal margin – <i>Go to question 336</i>
	335. Specify the liver size: centimeters
Labo	ratory studies at last evaluation prior to the start of the preparative regimen / infusion
336.	Date CBC drawn:
000.	YYYY MM DD
337.	WBC
	☐ Known – Go to question 338
	☐ Unknown – Go to question 339
	338
	$\Box \times 10^9 / L (\times 10^3 / mm^3)$
	□ x 10 ⁶ /L
339.	Neutrophils
	☐ Known – Go to question 340
	☐ Unknown – Go to question 341
	340%
341.	Blasts in blood
	☐ Known – Go to question 342
	□ Unknown – Go to question 343
	342 %
343.	Hemoglobin

CIBMTR Center Number:	CIBMTR Research ID:
☐ Known – Go to question 344	
☐ Unknown – Go to question 346	3
244	
344•	□ g/dL
	□ g/L
	□ mmol/L
345. Were RBCs transfused ≤ 30	days before date of test?
□ Yes	
□ No	
346. Platelets	
☐ Known – Go to question 347	
☐ Unknown – Go to question 34	19
247	
347	$ □ x 10^9 / L (x 10^3 / mm^3) $
	□ x 10 ⁶ /L
348. Were platelets transfused ≤	7 days before date of test?
□ Yes	
□ No	
349. Blasts in bone marrow	
☐ Known – Go to question 350	
☐ Unknown – Go to question 35	51
350 %	
351. Were tests for driver mutations perfo	ormed?
☐ Yes – Go to question 352	
□ No – Go to question 362	20
☐ Unknown – Go to question 36	12
352. JAK2	
☐ Positive – Go to question	on 353
☐ Negative – <i>Go to questi</i>	
☐ Not done – Go to quest	ion 355

CIBMTR Center Nun	nber: CIBMTR Research ID:
	353. JAK2 V617F
	□ Positive
	□ Negative
	□ Not done
	354. JAK2 Exon 12
	□ Positive
	☐ Negative
	☐ Not done
355.	CALR
	□ Positive – Go to question 356
	□ Negative – Go to question 359
	□ Not done – Go to question 359
	356. CALR type 1
	□ Positive
	□ Negative
	□ Not done
	357. CALR type 2
	□ Positive
	□ Negative
	□ Not done
	358. Not defined
	□ Positive
	□ Negative
	□ Not done
359.	MPL
	□ Positive
	□ Negative
	□ Not done
360.	CSF3R
	□ Positive
	□ Negative
	□ Not done

CIBMTR Center Number:	CIBMTR Research ID:
361. Was docume	entation submitted to the CIBMTR?
☐ Yes	
□ No	
362. Were cytogenetics to	ested (karyotyping or FISH)?
☐ Yes – Go to qu	estion 363
□ No – Go to que	estion 379
☐ Unknown – Go	to question 379
363. Were cytoge	netics tested via FISH?
□ Yes – Go	to question 364
□ No – Go t	to question 371
364. Sampl	e source
□ Blo	od
□ Bor	ne marrow
365. Result	s of tests
□ Abr	normalities identified – <i>Go to question</i> 366
□ No	abnormalities – Go to question 370
	cytogenetic abnormalities identified via FISH at last evaluation prior to the star preparative regimen / infusion
366.	International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
367.	Specify number of distinct cytogenetic abnormalities
	□ One (1)
	□ Two (2)
	☐ Three (3)
	☐ Four or more (4 or more)
368.	Specify abnormalities (check all that apply)
Мо	nosomy_
	□ <i>-</i> 5
	□ -7
	□ -Y

CIBMTR Center Number:	CIBMTR Research ID:
	+8
	+9
Translo	cation t(1;any)
	t(3q21;any)
	t(11q23;any)
	t(12p11.2;any)
	t(6;9)
Deletion	
	del(5q) / 5q-
	del(7q) / 7q-
	del(11q) / 11q-
	del(12p) / 12p-
	del(13q) / 13q-
	del(20q) / 20q-
Inversio	n
	dup(1)
	inv(3)
Other	
	i17q
	Other abnormality – <i>Go to question 369</i>
369.	Specify other abnormality:
370. Was docum	entation submitted to the CIBMTR? (e.g. FISH report)
☐ Yes	
□ No	
371. Were cytogenetics tested	via karyotyping?
☐ Yes – Go to question	
□ No – Go to question	1 379
372. Sample source	
☐ Blood	
☐ Bone marrow	
_ Solio mailow	

373. Results of tests

CIBMTR Center Number:	CIBMTR Research ID:
□ Ab	onormalities identified – <i>Go to question 374</i>
□ No	o evaluable metaphases – <i>Go to question 378</i>
□ No	o abnormalities – <i>Go to question</i> 378
	ify cytogenetic abnormalities identified via conventional cytogenetics at last evaluation r to the start of the preparative regimen / infusion
374.	International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
375.	Specify number of distinct cytogenetic abnormalities
	☐ One (1)
	□ Two (2)
	☐ Three (3)
	☐ Four or more (4 or more)
376.	Specify abnormalities (check all that apply)
Mono	osomy
	□ <i>-</i> 5
	□ - 7
	□ - Y
Triso	my □ +8
	□ +9
	□ 1 9
Trans	slocation □ t(1;any)
	□ t(3q21;any)
	□ t(11q23;any)
	□ t(12p11.2;any)
	□ t(6;9)
Delet	ion
	□ del(5q) / 5q-
	□ del(7q) / 7q-
	□ del(11q) / 11q-
	□ del(12p) / 12p-
	□ del(13q) / 13q-
	□ del(20q) / 20q-

Inversion	
□ dup(1)	
□ inv(3)	
Other	
□ i17q	
☐ Other abnormality – <i>Go to question 377</i>	
377. Specify other abnormality:	
378. Was documentation submitted to the CIBMTR? (e.g. karyotyping report)	
□ Yes	
□ No	
Status at transplantation / infusion	
379. What was the disease status?	
☐ Complete clinical remission (CR) – <i>Go to question 383</i>	
□ Partial clinical remission (PR) – <i>Go to question 383</i>	
☐ Clinical improvement (CI) – Go to question 380	
☐ Stable disease (SD) – Go to question 383	
☐ Progressive disease – Go to question 383	
☐ Relapse – Go to question 383	
□ Not assessed – Go to question 384	
380. Was an anemia response achieved?	
□ Yes	
□ No	
381. Was a spleen response achieved?	
□ Yes	
□ No	
382. Was a symptom response achieved?	
□ Yes	
□ No	
383. Date assessed:	

CIBMTR Cen	iter I	Number: CIBMTR Research ID:		
ı		Complete response (CR): Eradication of pre-existing abnormality – Go to question 385		
I		Partial response (PR): ≥ 50% reduction in abnormal metaphases – Go to question 385		
ı	☐ Re-emergence of pre-existing cytogenetic abnormality – <i>Go to question 385</i>			
!		Not assessed – Go to question 386		
1	☐ Not applicable – <i>Go to question 386</i>			
I		None of the above: Does not meet the CR or PR criteria – Go to question 385		
	385	5. Date assessed:		
386.	Spe	cify the molecular response		
1		Complete response (CR): Eradication of pre-existing abnormality – Go to question 387		
I		Partial response (PR): ≥50% decrease in allele burden – Go to question 387		
I		Re-emergence of a pre-existing molecular abnormality – <i>Go to question</i> 387		
I		Not assessed – Go to end of form		
ı		Not applicable – <i>Go to end of form</i>		
1		None of the above: Does not meet the CR or PR criteria – Go to question 387		
Others Levelse		7. Date assessed:		
Other Leuke	mıa	(OL)		
388. \$	Spe	cify the other leukemia classification		
		ure B-cell neoplasms Chronic lymphocytic leukemia (CLL), NOS (34) – <i>Go to question</i> 390		
1		Chronic lymphocytic leukemia / small lymphocytic lymphoma (71) – <i>Go to question 390</i>		
_		enic B-cell lymphomas and leukemias Hairy cell leukemia (35) – <i>Go to question 393</i>		
I		Splenic B-cell lymphoma / leukemia with prominent nucleoli (75) – <i>Go to question 393</i>		
I		T-prolymphocytic leukemia (74) – <i>Go to question</i> 390		
ı		Other leukemia, NOS (30) – <i>Go to question</i> 393		
I		Other leukemia (39) – Go to question 389		
	389	D. Specify other leukemia: – Go to question 393		
	390	0. Was any 17p abnormality detected?		
		☐ Yes – If disease classification is CLL, go to question 391. If PLL, go to question 393		
		□ No		

CIBMTR Cente	er Nui	mber: CIBMTR Research ID:
3	891.	Did a histologic transformation to diffuse large B-cell lymphoma (Richter syndrome) occur at any time after CLL diagnosis?
		☐ Yes – Go to question 395
		□ No – Go to question 393
St	atus	at transplantation / infusion
392. W	hat w	vas the disease status? (Atypical CML)
	Pri	mary induction failure – <i>Go to question</i> 394
	1st	complete remission (no previous bone marrow or extramedullary relapse) – Go to question 394
	2n	d complete remission – <i>Go to question 394</i>
	≥ 3	ord complete remission – Go to question 394
	1st	relapse – Go to question 394
	2n	d relapse – Go to question 394
	≥ 3	rd relapse – Go to question 394
	No	treatment – Go to end of form
3	393.	What was the disease status? (CLL, PLL, Hairy cell leukemia, Other leukemia)
		□ Complete remission (CR) – Go to question 394
		□ Partial remission (PR) – Go to question 394
		☐ Stable disease (SD) – Go to question 394
		□ Progressive disease (Prog) – Go to question 394
		☐ Untreated – Go to question 394
		□ Not assessed – Go to end of form
		394. Date assessed:
Hodgkin and N	Non-l	Hodgkin Lymphoma
395. Sp	pecify	the lymphoma histology <i>(at infusion)</i>
Ho	_	in Lymphoma assic Hodgkin lymphoma(150)
	Lyı	mphocyte depleted (154)
	Lyı	mphocyte-rich (151)
	Mix	xed cellularity (153)
	No	dular lymphocyte predominant Hodgkin lymphoma (155)
	No	dular sclerosis (152)

CIBMTR Center	Number: CIBMTR Research ID:
Bu	rkitt lymphoma Burkitt lymphoma (111)
La: □	rge B-cell lymphomas ALK-positive large B-cell lymphoma (1833)
	Diffuse, large B-cell lymphoma - Activated B-cell type subtype (1821) - Go to question 397
	Diffuse large B-cell lymphoma associated with chronic inflammation (1825)
	Diffuse, large B-cell lymphoma - Germinal center B-cell subtype (1820) - <i>Go to question</i> 397
	Diffuse large B-cell lymphoma / high-grade B-cell lymphoma with MYC and BCL2 rearrangements (1831)
	Diffuse large B-cell lymphoma/ high grade B-cell lymphoma with MYC and BCL6 rearrangements (1837)
	Diffuse large B-cell lymphoma/ high grade B-cell lymphoma with MYC, BCL2, and BCL6 rearrangements (1838)
	Diffuse large B-cell lymphoma, NOS (107)
	EBV-positive diffuse large B-cell lymphoma (1823)
	Fibrin-associated large B-cell lymphoma (1839)
	Fluid overload-associated large B-cell lymphoma (1840)
	High-grade B-cell lymphoma with 11q aberration (1834)
	Intravascular large B-cell lymphoma (136)
	Large B-cell lymphoma with IRF4 rearrangement (1832)
	Lymphomatoid granulomatosis (1835)
	Mediastinal grey zone lymphoma (149)
	Plasmablastic lymphoma (1836)
	Primary cutaneous diffuse large B-cell lymphoma, leg type (1822)
	Primary mediastinal large B-cell lymphoma (125)
	T-cell / histiocytic-rich large B-cell lymphoma (120)
	High-grade B-cell lymphoma, NOS (1830)
Pri	mary large B-cell lymphoma of immune-privileged sites Primary large B-cell lymphoma of the CNS (118)
	Primary large B-cell lymphoma of the testis (1881)
	Primary large B-cell lymphoma of the vitreoretina (1882)
KS	HV / HHV8-associated B-cell lymphoid proliferations and lymphomas KSHV / HHV8-positive diffuse large B-cell lymphoma (1826)
	Primary effusion lymphoma (138)
Ly	mphoplasmacytic lymphoma Lymphoplasmacytic lymphoma (173)
П	IdM-LPL / Waldenstrom macroglobulinemia (1883)

CIBMTR Center	r Number: CIBMTR Research ID:
	Non-IgM-LPL / Waldenstrom macroglobulinemia (1884)
Ma □	erginal zone lymphoma Extranodal marginal zone lymphoma of mucosa-associated lymphoid tissue (122)
	Nodal marginal zone lymphoma (123)
	Pediatric marginal zone lymphoma (1813)
	Primary cutaneous marginal zone lymphoma (1885)
Sp □	lenic B-cell lymphomas Splenic, B-cell lymphoma/leukemia with prominent nucleoli (1811)
	Splenic diffuse red pulp small B-cell lymphoma (1812)
	Splenic marginal zone lymphoma (124)
Fo	llicular lymphoma Duodenal-type follicular lymphoma (1815)
	Follicular, mixed, small cleaved and large cell (Grade II follicle center lymphoma) (103)
	Follicular, predominantly large cell (Grade IIIA follicle center lymphoma) (162)
	Follicular, predominantly large cell (Grade IIIB follicle center lymphoma) (163)
	Follicular, predominantly large cell (Grade IIIA vs IIIB not specified) (1814)
	Follicular, predominantly small cleaved cell (Grade I follicle center lymphoma) (102)
	Pediatric-type follicular lymphoma (1816)
	Follicular (grade unknown) (164)
Cu	taneous follicle center lymphoma Primary cutaneous follicle center lymphoma (1817)
Ma □	ntle cell lymphoma Mantle cell lymphoma (115)
	Leukemic non-nodal mantle cell lymphoma (1886)
Tra	Ansformations of indolent B-cell lymphomas Transformations of indolent B-cell lymphomas (1887)
Ly	mphomas associated with immune deficiency and dysregulation Classical Hodgkin lymphoma PTLD (1876)
	EBV-positive mucocutaneous ulcer (1824)
	Hyperplasia arising in immune deficiencies (e.g. PTLD) (1871)
	Infectious mononucleosis PTLD (1872)
	Monomorphic PTLD (B- and T-/NK-cell types) (1875)
	Polymorphic lymphoproliferative disorders arising in immune deficiency/dysregulation (1874)

CIBMTR Center	Number: CIBMTR Research ID:
	Adult T-cell lymphoma / leukemia (134)
	Aggressive NK-cell leukemia (27)
	NK-large granular lymphocytic leukemia (1856)
	Sézary syndrome (142)
	T-large granular lymphocytic leukemia (126)
Prii	mary cutaneous T-cell lymphomas Mycosis fungoides (141)
	Primary cutaneous acral CD8-positive lymphoproliferative disorder (1853)
	Primary cutaneous CD4-positive small or medium T-cell lymphoproliferative disorder (1854)
	Primary cutaneous CD8-positive aggressive epidermotropic cytotoxic T-cell lymphoma (1852)
	Primary cutaneous CD30-positive T-cell lymphoproliferative disorder: lymphomatoid papulosis (147)
	Primary cutaneous CD30-positive T-cell lymphoproliferative disorder: primary cutaneous anaplastic large cell lymphoma (1888)
	Primary cutaneous gamma / delta T-cell lymphoma (1851)
	Subcutaneous panniculitis-like T-cell lymphoma (146)
	Primary cutaneous peripheral T-cell lymphoma, NOS (1889)
Inte	estinal T-cell and NK-cell lymphoid proliferations and lymphomas Enteropathy-associated T-cell lymphoma (133)
	Indolent T-cell lymphoma of the gastrointestinal tract (1858)
	Indolent NK-cell lymphoproliferative disorder of the gastrointestinal tract (1890)
	Monomorphic epitheliotropic intestinal T-cell lymphoma (1857)
	Intestinal T-cell lymphoma, NOS (1891)
	Datosplenic T-cell lymphoma Hepatosplenic T-cell lymphoma (145)
Ana	aplastic large cell lymphoma ALK-positive anaplastic large cell lymphoma (143)
	ALK-negative anaplastic large-cell lymphoma (144)
	Breast implant–associated anaplastic large cell lymphoma (1861)
No.	dal T-follicular helper (TFH) cell lymphoma Nodal TFH cell lymphoma, angioimmunoblastic-type (131)
	Nodal TFH cell lymphoma, follicular-type (1859)
	Nodal TFH cell lymphoma, NOS (1860)
	ner peripheral T-cell lymphomas Peripheral T-cell lymphoma, NOS (130)

CIBMTR Ce	nter N	umber: CIBMTR Research ID:
		positive NK/T-cell lymphomas BV-positive nodal T- and NK-cell lymphoma (1892)
		xtranodal NK / T-cell lymphoma (137)
		positive T- and NK-cell lymphoid proliferations and lymphomas of childhood systemic EBV-positive T-cell lymphoma of childhood (1855)
		other B-cell lymphoma (129) – <i>Go to question</i> 396
		Other T-cell / NK-cell lymphoma (139) – <i>Go to question</i> 396
	396.	Specify other lymphoma histology:
	397.	Assignment of DLBCL (germinal center B-cell type vs. activated B-cell type) subtype was based or
		☐ Immunohistochemistry (e.g. Han's algorithm)
		☐ Gene expression profile
		☐ Unknown method
398.	Is the	lymphoma histology reported at transplant a transformation from CLL?
	□ Y	es – Also complete Chronic Lymphocytic Leukemia (CLL) Form 2013 – Go to question 399
		o – Go to question 400
	399.	Was any 17p abnormality detected?
		☐ Yes – Go to question 404
		□ No – Go to question 404
400.		lymphoma histology reported at transplant a transformation from a different lymphoma histology?
	□ Y	es – Go to question 401
		o – Go to question 404
	401.	Specify the original lymphoma histology (prior to transformation):
		402. Specify other lymphoma histology:
	403.	Date of original lymphoma diagnosis:
404.	Was a	a PET (or PET/CT) scan performed? (at last evaluation prior to the start of the preparative regimen /
	□ Y	es – Go to question 405
		o – Go to question 410

405. Was the PET (or PET/CT) scan positive for lymphoma involvement at any disease site?

CIBMTR Cer	nter	lumber: CIBMTF	Research ID:		
		☐ Yes			
		□ No			
	40	Date of PET (or PET/CT) scan			
		☐ Known – Go to question 407			
		☐ Unknown – Go to question 408			
		407. Date of PET (or PET/CT) scan: _		· —	·
			YYYY	MM	DD
	40	. Deauville (five-point) score of the PET	or PET/CT) scan		
		☐ Known – Go to question 409			
		☐ Unknown – Go to question 410			
		409. Scale			
		☐ 1- no uptake or no residual up	itake		
		☐ 2- slight uptake, but below blo		um)	
		☐ 3- uptake above mediastinal,			the liver
		☐ 4- uptake slightly to moderate	-	•	
		☐ 5- markedly increased uptake			
	Sta	us at transplantation / infusion			
410.	Wh	t was the disease status?			
		Disease untreated – Go to end of form			
		PIF res - Primary induction failure – resista progressive disease on treatment. – Go to		IPLETE ren	nission but with stable or
		PIF sen / PR1 - Primary induction failure – remission on treatment. – <i>Go to question</i>		in COMPLE	TE remission but with partial
		PIF unk - Primary induction failure – sensi	ivity unknown – Go	to questi	on 411
		CR1 - 1 st complete remission: no bone ma question 411	rrow or extramedul	lary relapse	prior to transplant – <i>Go to</i>
		CR2 - 2 nd complete remission – Go to que	estion 411		
		CR3+ - 3 rd or subsequent complete remiss	ion – Go to quest	ion 411	
		REL1 unt - 1 st relapse – untreated; include question 411	s either bone marro	ow or extran	nedullary relapse – Go to
		REL1 res - 1 st relapse – resistant: stable o	progressive disea	se with trea	tment – Go to question 411
		REL1 sen - 1 st relapse – sensitive: partial ı CR2) – Go to question 411	emission (if comple	ete remissio	n was achieved, classify as
		REL1 unk - 1 st relapse – sensitivity unknov	/n – Go to questi o	on 411	

CIBMTR Center	Number: CIBMTR Research ID:
	REL2 unt - 2 nd relapse – untreated: includes either bone marrow or extramedullary relapse – <i>Go to question 411</i>
	REL2 res - 2 nd relapse – resistant: stable or progressive disease with treatment – <i>Go to question 411</i>
	REL2 sen - 2^{nd} relapse – sensitive: partial remission (if complete remission achieved, classify as CR3+) – <i>Go to question 411</i>
	REL2 unk - 2 nd relapse – sensitivity unknown – <i>Go to question 411</i>
	REL3+ unt - 3rd or subsequent relapse – untreated; includes either bone marrow or extramedullary relapse – <i>Go to question 411</i>
	REL3+ res - 3^{rd} or subsequent relapse – resistant: stable or progressive disease with treatment – \textbf{Go} to $\textbf{question 411}$
	REL3+ sen - 3^{rd} or subsequent relapse – sensitive: partial remission (if complete remission achieved, classify as CR3+) – Go to question 411
	REL3+ unk - 3 rd relapse or greater – sensitivity unknown – <i>Go to question 411</i>
41	Total number of lines of therapy received (between diagnosis and HCT / infusion)
	□ 1 line
	□ 2 lines
	□ 3+ lines
	412. Date assessed:
	YYYY MM DD
Multiple Myelor	na / Plasma Cell Disorder (PCD)
413 Sne	cify the multiple myeloma/plasma cell disorder (PCD) classification
	Multiple myeloma (178) – <i>Go to question 415</i>
_	Multiple myeloma-light chain only (186) – <i>Go to question 415</i>
_	Multiple myeloma-non-secretory (187) – <i>Go to question 421</i>
	Plasma cell leukemia (172) – <i>Go to question 423</i>
_	Plasmacytoma (175) – <i>Go to question 420</i>
	Smoldering myeloma (180) – <i>Go to question 423</i>
	Immuno-globulin-related (AL) amyloidosis (174) – <i>Go to question 416</i>
Pla □	sma cell neoplasm with associated paraneoplastic syndrome POEMS syndrome (176) – Go to question 423
	Monoclonal gammopathy of renal significance (MGRS) (1611) - Go to question 417
	Other plasma cell disorder (179) – Go to question 414
41	4. Specify other plasma cell disorder: – Go to question 423
41	5. Specify heavy and/or light chain type <i>(check all that apply)</i>

CIBMTR Center Nu	mber: CIBMTR Research ID:
	☐ IgG kappa – Go to question 421
	☐ IgA kappa – Go to question 421
	☐ IgM kappa – Go to question 421
	☐ IgD kappa – Go to question 421
	☐ IgE kappa – Go to question 421
	☐ IgG lambda – <i>Go to question 421</i>
	□ IgA lambda – <i>Go to question 421</i>
	☐ IgM lambda – <i>Go to question 421</i>
	□ IgD lambda – <i>Go to question 421</i>
	☐ IgE lambda – <i>Go to question 421</i>
	☐ IgG (heavy chain only) – <i>Go to question 421</i>
	☐ IgA (heavy chain only) – Go to question 421
	☐ IgM (heavy chain only) – <i>Go to question 421</i>
	☐ IgD (heavy chain only) – Go to question 421
	☐ IgE (heavy chain only) – Go to question 421
	☐ Kappa (light chain only) – Go to question 421
	□ Lambda (light chain only) – <i>Go to question 421</i>
416.	Specify Amyloidosis classification
	☐ AL amyloidosis – <i>Go to question 423</i>
	□ AH amyloidosis – <i>Go to question 423</i>
	□ AHL amyloidosis – <i>Go to question 423</i>
417.	Select monoclonal gammopathy of renal significance (MGRS) classification
	☐ Light chain Fanconi syndrome – <i>Go to question 419</i>
	☐ Proximal tubulopathy without crystals – <i>Go to question 419</i>
	☐ Crystal-storing histiocytosis – Go to question 419
	□ Non-amyloid fibrillary glomerulonephritis – <i>Go to question 419</i>
	☐ Immunotactoid glomerulopathy (ITGN)/ Glomerulonephritis with organized monoclonal microtubular immunoglobulin deposits (GOMMID) – <i>Go to question 419</i>
	☐ Type 1 cryoglobulinemic glomerulonephritis – <i>Go to question 419</i>
	☐ Monoclonal immunoglobulin deposition disease (MIDD) – <i>Go to question 418</i>
	□ Proliferative glomerulonephritis with monoclonal immunoglobulin G deposits (PGNMID) – Go to question 419
	☐ C3 glomerulopathy with monoclonal gammopathy – <i>Go to question 419</i>
	☐ Unknown – Go to question 419

418. Select monoclonal immunoglobulin deposition disease (MIDD) subtype

CIBMTR Ce	enter	Number:	CIBMTR Research ID:
			☐ Light chain deposition disease (LCDD)
			☐ Monoclonal immunoglobulin deposition disease
			☐ Heavy chain deposition disease (HCDD)
		419.	Was documentation submitted to the CIBMTR? (e.g. pathology report)
			☐ Yes – Go to question 423
			□ No – Go to question 423
	42	20. Solita	ry plasmacytoma was
		□Ех	traosseous plasmacytoma – <i>Go to question 423</i>
		□ Sc	olitary plasmacytoma of bone – Go to question 423
421.			e Durie-Salmon staging? (at diagnosis, or if subsequent infusion, report based on last relapse prior to this infusion)
		bone stru	All of the following: Hgb > 10g/dL; serum calcium normal or <10.5 mg/dL; bone x-ray normal acture (scale 0), or solitary bone plasmacytoma only; low M-component production rates IgG IgA < 3g/dL; urine light chain M-component on electrophoresis <4g/24h) — Go to question
		Stage II	(Fitting neither Stage I or Stage III) – Go to question 422
		bone les	(One of more of the following: Hgb < 8.5 g/dL; serum calcium > 12 mg/dL; advanced lytic ions (scale 3); high M-component production rates IgG >7g/dL, IgA > 5g/dL; Bence Jones 12g/24h) – Go to question 422
		Unknowr	n – Go to question 423
	42		was the Durie-Salmon sub classification? (at diagnosis, or if subsequent infusion, report d on last relapse / progression prior to this infusion)
		□ A ·	- relatively normal renal function (serum creatinine < 2.0 mg/dL)
		□ B ·	- abnormal renal function (serum creatinine ≥ 2.0 mg/dL)
423.	Did	I the recipi	ient have a preceding or concurrent plasma cell disorder?
		Yes – G	o to question 424
		No – Go	to question 427
	Со	py questi	ons 424-426 to report more than one concurrent or preceding disorder
	42	24. Speci	fy preceding / concurrent disorder
		□М	ultiple myeloma – <i>Go to question 426</i>
		□Мι	ultiple myeloma-light chain only – Go to question 426
		□М	ultiple myeloma-non-secretory – Go to question 426
		□ Pla	asma cell leukemia – <i>Go to question 426</i>
		□ Pla	asmacytoma – Go to question 426

CIBMTR Center	Number: ₋			CIBMTR	Research ID	:								
	□ Sm	☐ Smoldering myeloma – <i>Go to question 426</i> ☐ Immuno-globulin-related (AL) amyloidosis – <i>Go to question 426</i>												
	□ Imi													
		Plasma cell neoplasm with associated paraneoplastic syndrome □ POEMS syndrome − Go to question 426 □ Monoclonal gammopathy of unknown significance (MGUS) − Go to question 426 □ Monoclonal gammopathy of renal significance (MGRS) − Go to question 426 □ Other plasma cell disorder (PCD) − Go to question 425												
	□Мо													
	□Мо													
	□ Otl													
	425.	Specify other	r preceding/	/concurren	t disorder: _									
	426.	Date of diagn	nosis of pred	ceding / co	oncurrent dis	order:		- <u></u> -						
							YYYY	MM	DD					
Сор	y questic	ons 424-426 t	to report m	ore than	one concur	rent or pre	eceding disord	der						
Lab infusion	s at diag	nosis, <i>or if</i> su	ubsequent	infusion,	report base	ed on last	relapse / prog	ression pric	or to this					
427. Ser	rum β2-microglobulin													
	Known –	Go to quest	ion 428											
	Unknown	n – Go to que	estion 429											
42	8. Serum	n β2-microglol	bulin:	•										
						□ µg/d	dL							
						□ mg/	L							
						□ nmo	ol/L							
429. Sert	um album	iin												
	Known –	Go to quest	ion 430											
	Unknown	n – Go to que	estion 431											
43	0. Serum	n albumin:	•	_										
				□ g/dL										
				□ g/L										
I.S.S this infus	_	nosis, or if s	ubsequent	t infusion,	report base	ed on last	relapse / prog	ression pri	or to					
431. Staç	ge													
	Known –	Go to quest	ion 432											
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CIBMTR Center No	umber: CIBMTR Research ID:
432.	Stage
	□ 1 (Serum β2-microglobulin < 3.5 mg/L, Serum albumin ≥ 3.5 g/dL)
	□ 2 (Not fitting stage 1 or 3)
	□ 3 (Serum β2-microglobulin ≥ 5.5 mg/L; Serum albumin —)
R - I.S this infusio	S.S. at diagnosis, or if subsequent infusion, report based on last relapse / progression prior to n
433. Stage	
□К	nown – Go to question 434
□ U	nknown – Go to question 435
434.	Stage
	□ 1 (ISS stage I and no high-risk cytogenetic abnormalities by FISH [deletion 17p / 17p-, t(4;14), t(14;16)] and normal LDH levels)
	□ 2 (Not R-ISS stage I or III)
	□ 3 (ISS stage III and either high-risk cytogenetic abnormalities by FISH [deletion 17p / 17p-, t(4;14), t(14;16)] or high LDH levels)
Labs at d infusion	iagnosis, or if subsequent infusion, report based on last relapse / progression prior to this
435. Plasm	na cells in peripheral blood by flow cytometry
□К	nown – Go to question 436
□ U	nknown – Go to question 437
436.	• %
437. Plasm	na cells in peripheral blood by morphologic assessment
□К	nown – Go to question 438
□ U	nknown – Go to question 440
438.	%
439.	·
	$\Box \times 10^{9}/L (\times 10^{3}/mm^{3})$
	□ x 10 ⁶ /L
440. LDH	
□К	nown – Go to question 441
□ U	nknown – Go to question 443

CIBMTR Center Number:	CIBMTR Research ID:
441	•_
	□ U/L
	□ µkat/L
442. Upper limit	of normal for LDH: •
Labs at diagnosis infusion	s, or if subsequent infusion, report based on last relapse / progression prior to this
	tested (karyotyping or FISH)? (at diagnosis, or if subsequent infusion, report based on pression prior to this infusion)
☐ Yes – Go to q	uestion 444
□ No – Go to q u	uestion 456
□ Unknown – G o	o to question 456
444. Were cytog	genetics tested via FISH?
□ Yes – G	o to question 445
□ No – G o	to question 450
445. Resu	ults of tests
□А	bnormalities identified – <i>Go to question 446</i>
□N	o abnormalities – Go to question 449
446	. International System for Human Cytogenetic Nomenclature (ISCN) compatible string:
447	. Specify abnormalities <i>(check all that apply)</i>
	Trisomy □ +3
	□ +5
	□ +7
	□ +9
	□ +11
	□ +15
	□ +19
	Translocation □ t(4;14)
	□ t(6;14)
	□ t(11;14)

CIBMTR Center Number:		CIBMTR Research ID:					
		t(14;16)					
		t(14;20)					
	Delet	ion					
		del (13q) / 13q-					
		del (17p) / 17p-					
		osomy - 13					
		- 17					
	Othe	r Hyperdiploid (>50)					
		Hypodiploid (<46)					
		MYC rearrangement					
		Any abnormality at 1q					
		Any abnormality at 1p					
		Other abnormality – <i>Go to question 448</i>					
	448.	Specify other abnormality:					
449	9. Was docum	entation submitted to the CIBMTR? (e.g. FISH report)					
	☐ Yes						
	□ No						
450. Wei	re cytogenetics	s tested via karyotyping?					
	res – Go to q	uestion 451					
1 🗆	No – Go to qu	estion 456					
451	I. Results of to	ests					
		alities identified – Go to question 452					
		rable metaphases – Go to question 455					
		rmalities – Go to question 455					
	452. Inter	rnational System for Human Cytogenetic Nomenclature (ISCN) compatible string:					
	453. Spe	cify abnormalities (check all that apply)					
	Triso	my +3					
	п	+5					

CIBMTR Center Number:	CIBMTR Research ID:
	+7
	+9
	+11
	+15
	+19
	slocation t(4;14)
	t(6;14)
	t(11;14)
	t(14;16)
	t(14;20)
Delet	tion del (13q) / 13q-
	del (17p) / 17p-
	uei (17 <i>p)</i> / 17 <i>p</i> -
	osomy
	- 13
Ц	- 17
Othe	r
	Hyperdiploid (>50)
	Hypodiploid (<46)
	MYC rearrangement
	Any abnormality at 1q
	Any abnormality at 1p
	Other abnormality – Go to question 454
454.	Specify other abnormality:
455. Was docum	nentation submitted to the CIBMTR? (e.g. karyotyping report)
☐ Yes	
□ No	
Ctatus at transmigntation	a / infusion
Status at transplantation	17 musion
456. What is the hematologic of	lisease status?
☐ Stringent complete re	sponse (sCR)
☐ Complete response (CR)
☐ Very good partial resp	ponse (VGPR)

CIBMTR Ce	nter	Number: C	IBMTR Research ID:
		Partial response (PR)	
		No response (NR) / stable disease (SD)
		Progressive disease (PD)	
		Relapse from CR (Rel) (untreated)	
		Unknown	
	45	77. Date assessed:	
458.	Spe	ecify amyloidosis hematologic respon	se (for Amyloid patients only)
		Complete response (CR)	
		Very good partial response (VGPR)	
		Partial response (PR)	
		No response (NR) / stable disease (SD)
		Progressive disease (PD)	
		Relapse from CR (Rel) (untreated)	
		Unknown	
	45	9. Date assessed:	Go to end of form MM DD
Solid Tumo	rs		
460.	Spe	ecify the solid tumor classification	
		east cancer Breast cancer (250)	
	Tur	mors of the head / neck Tumors of the head / neck (201)	
	Dig □	cestive system tumors Colorectal (228)	
		Pancreatic (206)	
		Tumor of the esophagus and gastro	-esophageal (GE) junction (2111)
		Tumors of the stomach (229)	
		Tumors of liver and intrahepatic bile	ducts (207)
	Cei	ntral nervous system tumors Atypical teratoid rhabdoid tumor (AT	RT) (2212)
		Central nervous system tumor, inclu	ding CNS PNET (220)

CIBMTR Center	r Number: CIBMTR F	CIBMTR Research ID:				
	Diffuse intrinsic pontine glioma (DIPG) (2213)				
	Ependymoma (2214)					
	Glioblastoma multiforme (GBM) (2215)					
	Medulloblastoma (226)					
Sof	oft tissue or bone tumors Bone sarcoma (excluding Ewing family tumo	rs) (273)				
	Desmoplastic small round cell tumors (2216)					
	Ewing family tumors of bone (including PNE	Γ) (275)				
	Ewing family tumors, extraosseous (including	g PNET) (276)				
	Malignant Peripheral Nerve Sheath Tumor (2	248)				
	Myxoid round cell sarcoma (2217)					
	Rhabdomyosarcoma (232)					
	Synovial sarcoma (245)					
	Other soft tissue sarcoma (excluding Ewing	family tumors) (274)				
Tui	mors of endocrine organs Germ cell tumor, gonadal (2218)					
	Germ cell tumor, extragonadal (225)					
	Neuroblastoma (222)					
The	oracic tumors Adenocarcinoma (2219)					
	Lung, non-small cell (203)					
	Lung, small cell (202)					
	Lung, not otherwise specified (230)					
	Squamous carcinoma (2220)					
	Tumor of the pleura (Mesothelioma) (2221)					
Ski	in tumors Melanoma (219)					
Ge □	enitourinary tumors Ovarian (epithelial) (214)					
	Prostate (209)					
	Renal cell (208)					
	Testicular (210)					
	Vaginal (215)					
	ediatric-focused tumors	222)				

CIBMTR Center Number:		Number: CIBMTR Research ID:						
		Retinoblastoma (223)						
		Wilms tumor (221)						
	Oth	er solid tumors Solid tumor, not otherwise specified (200)						
		Other solid tumor (269) – <i>Go to question 461</i>						
	46	1. Specify other solid tumor: – Go to end of form						
Aplastic An	emi	a						
462.	-	ecify the aplastic anemia classification – If the recipient developed MDS or AML, indicate MDS or ML as the primary disease.						
		Acquired AA, not otherwise specified (301) - Go to question 463						
		Acquired AA secondary to chemotherapy (313) – Go to question 463						
		Acquired AA secondary to hepatitis (302) (any form of hepatitis) - Go to question 463						
	□ Acquired AA secondary to immunotherapy or immune effector cell therapy (314) – Go to quest 463							
☐ Acquired AA secondary to toxin / other drug (303) – Go to question 463								
	☐ Acquired amegakaryocytosis (not congenital) (304) – <i>Go to end of form</i>							
	☐ Acquired pure red cell aplasia (not congenital) (306) – Go to end of form							
		Other acquired cytopenic syndrome (309) – <i>Go to question 464</i>						
	46	3. Specify severity						
		☐ Severe / very severe – Go to end of form						
		□ Not severe – Go to end of form						
	46	4. Specify other acquired cytopenic syndrome: – Go to end of form						
Inherited Bo	one	Marrow Failure Syndromes						
465.	-	ecify the inherited bone marrow failure syndrome classification – If the recipient developed MDS or ML, indicate MDS or AML as the primary disease.						
		Diamond-Blackfan anemia (pure red cell aplasia) (312) – Go to end of form						
		Telomere Biology Disorders including Dyskeratosis congenita (DKC1, TERT, TERC, and other mutations) (307) $-$ Go to end of form						
		Fanconi anemia (311) – <i>Go to end of form</i>						
		Severe congenital neutropenia (Elastase deficiency / ELANE or Kostmann disease / HAX1 mutations) (460) – <i>Go to end of form</i>						
		Shwachman-Diamond (DNAJC21, EFL1, or SBDS mutations) (305) – Go to end of form						
		Germline <i>SAMD9</i> variant (MIRAGE Syndrome) (2311) – <i>Go to end of form</i>						

CIBMTR Cente	er Nu	umber: CIBMTR Research ID:
		ermline <i>SAMD9L</i> variant (SAMD9L-related Ataxia Pancytopenia Syndrome) (2312) – Go to end of
	O	ther inherited bone marrow failure syndrome (339) – Go to end of form
Hemoglobinop	path	ies
466. Sp		fy the hemoglobinopathy classification
		ickle cell disease (356) – Go to question 469
	Tr	ransfusion dependent thalassemia (360) – Go to question 467
	O	ther hemoglobinopathy (359) – Go to question 468
4	167.	Specify transfusion dependent thalassemia
		☐ Transfusion dependent beta thalassemia (357) – Go to question 469
		☐ Other transfusion dependent thalassemia (358) – <i>Go to question 469</i>
4	168.	Specify other hemoglobinopathy:
C	Ques	stions 469-501 are for sickle cell disease and transfusion dependent thalassemia
4	169.	Was tricuspid regurgitant jet velocity (TRJV) measured by echocardiography?
		☐ Yes – Go to question 470
		□ No – Go to question 472
		☐ Unknown – Go to question 472
		470. TRJV measurement
		☐ Known – Go to question 471
		☐ Unknown – Go to question 472
		471. TRJV measurement:
4	172.	Was liver iron content (LIC) tested within 6 months prior to infusion?
		☐ Yes – Go to question 473
		□ No – Go to question 475
		473. Liver iron content: •
		☐ mg Fe/g liver dry weight
		□ g Fe/kg liver dry weight
		□ µmol Fe/g liver dry weight

474. Method used to estimate LIC?

CIBMTR Center Nun	nber:		CIBMTR Research ID:						
		T2*MRI							
		SQUID M	1RI						
		FerriScar	า						
		Liver biop	psy						
		Other							
	Is the recipient red blood cell transfusion dependent? (requiring transfusion to maintain HGB 9 g/dL)								
	□ Yes –	Go to qu	uestion 476						
	□ No – (Go to qu	estion 483						
	476. Yea	ar of first	of first transfusion (since diagnosis):						
	477. Wa	as iron ch	elation therapy given at any time since diagnosis?						
		□ Yes – Go to question 478							
		No – Go	No – Go to question 483						
		Unknow	n – Go to question 483						
	47	first t	ron chelation therapy meet the following criteria: initiated within 18 months of the transfusion and administered for at least 5 days / week (either oral or parenteral chelation medication)?						
			Yes, iron chelation therapy given as specified – <i>Go to question 481</i>						
			No, iron chelation therapy given, but not meeting criteria listed – <i>Go to question 479</i>						
			Iron chelation therapy given, but details of administration unknown – <i>Go to question 481</i>						
		479.	Specify reason criteria not met						
			□ Non-adherence – <i>Go to question 481</i>						
			☐ Toxicity due to iron chelation therapy – <i>Go to question 481</i>						
			□ Other – Go to question 480						
			480. Specify other reason criteria not met:						
	48	31. Year	iron chelation therapy started						
			Known – Go to question 482						
			Unknown – Go to question 483						
		482.	Year started:						

CIBMTR Center Nu	ımber:				CI	IBMTR Rese	arch ID):				
483.	Did the	recipi	ent ha	ve hepat	ome	galy? <i>(</i> ≥ 2 <i>cn</i>	n below	costal m	nargin)			
	☐ Yes – Go to question 484											
	□ No – Go to question 485											
	□ Unkr	□ Unknown – Go to question 485										
	484. Liver size as measured below the costal margin at most recent evaluation:											
485.	Was a liver biopsy performed at any time since diagnosis?											
	☐ Yes – Go to question 486											
	□ No –	- Go t	o que	stion 49	93							
	486. D	Date as	ssesse	ed								
		□ Kno	wn – (Go to qι	ıesti	on 487						
		□ Unk	nown	– Go to	que	stion 488						
		487. Date assessed:					☐ Date estimated					
		407.	Duto	45505500	 -	YYYY		MM	DD	_ Date commuted		
	488. V	Vas th	ere ev	idence o	of live	r cirrhosis?						
		□ Yes										
		□ No										
		□ Unk	nown									
	400 \	A/ 4l-			£ 15	6 1						
						r fibrosis?						
				to ques								
		□ No – Go to question 491 □ Unknown – Go to question 491										
		LI OIIK	IIIOWIII	- G0 10	que	511011 491						
	4	490.	Туре	of fibrosi	s							
				Bridging								
				Periporta	ıl							
				Other								
				Unknowr	1							
	491. V	Vas th	ere ev	idence o	of chro	onic hepatitis	s?					
		□ Yes										
	Г	□ No										
		□ Unk	nown									

492. Was documentation submitted to the CIBMTR? (e.g., liver biopsy)

CIBMTR Center Nu	mber: CIBMTR Research ID:
	□ Yes
	□ No
493.	Is there evidence of abnormal cardiac iron deposition based on MRI of the heart at time of infusion?
	□ Yes
	□ No
494.	Did the recipient have a splenectomy?
	□ Yes
	□ No
	□ Unknown
Labo	ratory studies at last evaluation prior to start of preparative regimen
495.	Serum iron
	☐ Known – Go to questions 496
	☐ Unknown – Go to questions 497
	496. Serum iron: •
	□ μg/dL
	□ μmol/L
497.	Total iron binding capacity (TIBC)
	☐ Known – Go to question 498
	☐ Unknown – Go to question 499
	498. TIBC: •
	□ μg/dL
	□ µmol/L
400	Total common hillion him
499.	Total serum bilirubin
	□ Known – Go to question 500 □ Unknown – Go to end of form
	Unknown – Go to ena or form
	500. Total serum bilirubin:
	□ mg/dL
	□ μmol/L
	501. Upper limit of normal for total serum bilirubin: ●

CIBMTR Center Number:	CIBMTR Research ID:
OIDWITT OCHLOT HUMBOL.	OIDMITT TOSCATOLID.

Disorders of the Immune System

502. Specify disorder of immune system classification

Se	vere Combined Immunodeficiencies SCID, T- B+ NK-, JAK3 mutation (2411)– Go to question 506
	SCID, T- B+ NK-, IL2RG mutations, X-linked SCID (2412) - Go to question 506
	SCID, T- B- NK-, Adenosine deaminase (ADA) deficiency (401) – Go to question 506
	SCID, T- B- NK-, reticular dysgenesis (405) – <i>Go to question 506</i>
	SCID, T- B- NK+, RAG 1/2 deficiency (2413) - Go to question 506
	SCID, T- B- NK+, DCLRE1C (Artemis) deficiency (2414) - Go to question 506
	SCID, T- B+ NK+, ILR alpha deficiency (403) – Go to question 506
	SCID, T- B- NK-, NOS (2415) – Go to question 506
	SCID, not otherwise specified (410) – Go to question 506
	Other SCID (with known genetic mutation) (419) – <i>Go to question 503</i>
Co	mbined Immunodeficiencies CD40 ligand deficiency (464) – Go to question 506
	DOCK8 Deficiency (2416) – Go to question 506
	MHC Class II Deficiency (Bare lymphocyte syndrome) (406) – <i>Go to question 506</i>
	Omenn syndrome (404) – Go to question 506
	ZAP-70 deficiency (2417) – Go to question 506
Co	mbined Immunodeficiencies with Associated or Syndromic Features Ataxia telangiectasia (451) – Go to question 506
	Cartilage-hair hypoplasia (462) – <i>Go to question 506</i>
	DiGeorge anomaly (454) – Go to question 506
	NEMO Deficiency Syndrome (2418) – Go to question 506
	Wiskott-Aldrich syndrome (453) – <i>Go to question 506</i>
Pre	edominately Antibody deficiencies Common variable immunodeficiency (457) – Go to question 506
	Activated PI3 Kinase Delta Deficiency Syndrome (APDS1 or PIK3CD) (2419) - Go to question 506
Dis	ceases of immune dysregulation, hemophagocytic lymphohistiocytosis Chediak-Higashi syndrome (456) – Also complete Pigmentary Dilution Disorder (PDD) Pre-HCT Data Form – Go to question 506
	Griscelli syndrome type 2 (465) – Also complete Pigmentary Dilution Disorder (PDD) Pre-HCT Data Form – Go to question 506
	Hermansky-Pudlak syndrome type 2 (466) – Also complete Pigmentary Dilution Disorder (PDD) Pre-HCT Data Form – Go to question 506

CIBMTR Center	Number: C	IBMTR Research ID:		
		Other pigmentary dilution disorder (469) – Also complete Pigmentary Dilution Disorder (PDD) Pre- HCT Data Form – Go to question 505		
Dis □	seases of immune dysregulation, E SAP deficiency (XIAP-1) (458) – Go			
	XIAP-2 deficiency (2420) – Go to q	uestion 506		
	ITK deficiency (2421) – Go to ques	stion 506		
Dis □		yndromes with Autoimmunity and Others, NOS yndrome (ALPS) (2422) – Go to question 506		
	CTLA4 deficiency (2423) - Go to q	uestion 506		
	IPEX, Immune Dysregulation Polyer <i>Go to question 506</i>	ndocrinopathy, enteropathy X-linked (FOXP3 deficiency) (2424) –		
	LRBA Deficiency (2425) - Go to qu	uestion 506		
	STAT3 Gain of Function (2426) – G	o to question 506		
Co	ngenital defects of phagocyte Chronic granulomatous disease (45	5) – Go to question 506		
	GATA2 deficiency (2427) - Go to q	27) – Go to question 506		
	Leukocyte adhesion deficiencies (45	59) – Go to question 506		
	Neutropenia with combined immune question 506	e deficiency (MKL1 deficiency, Actin deficiency) (461) – <i>Go to</i>		
Oth	ner Immunodeficiencies HIV infection (452) – Go to questic	on 506		
	STAT1 Gain of Function (2428) - G	o to question 506		
	Other immunodeficiencies (479) – G	Go to question 504		
	Immune deficiency, not otherwise sp	pecified (400) - Go to question 506		
50	03. Specify other SCID:	– Go to question 506		
50	04. Specify other immunodeficiency:	Go to question 506		
50	05. Specify other pigmentary dilution 506	disorder: – Go to question		
50	06. Did the recipient have an active o	or recent infection with a viral pathogen within 60 days of HCT?		
	☐ Yes – Go to question 507			
	□ No – Go to question 508			
	507. Specify viral pathogen (ch	eck all that apply)		
	☐ 304 Adenovirus			

CIBMTR Center Numbe	r: CIBMTR Research ID:	
	□ 341 BK Virus	
	□ 344 Coronavirus	
□ 303 Cytomegalovirus (CMV)		
☐ 347 Chikungunya Virus ☐ 346 Dengue Virus		
	□ 327 Enterovirus D68 (EV-D68)	
	□ 326 Enterovirus (polio)	
	□ 328 Enterovirus NOS	
	□ 318 Epstein-Barr Virus (EBV)	
	□ 306 Hepatitis A Virus	
	□ 307 Hepatitis B Virus	
	□ 308 Hepatitis C Virus	
	□ 340 Hepatitis E	
	□ 301 Herpes Simplex Virus (HSV)	
☐ 317 Human herpesvirus 6 (HHV-6)		
□ 309 Human Immunodeficiency Virus 1 or 2		
	□ 343 Human metapneumovirus	
	□ 322 Human Papillomavirus (HPV)	
	□ 349 Human T-lymphotropic Virus 1 or 2	
	□ 310 Influenza, NOS	
	□ 323 Influenza A Virus	
	□ 324 Influenza B Virus	
	☐ 342 JC Virus (Progressive Multifocal Leukoencephalopathy (PML))	
	□ 311 Measles Virus (Rubeola)	
	□ 312 Mumps Virus	
	□ 345 Norovirus	
	□ 316 Human Parainfluenza Virus (all species)	
	☐ 314 Respiratory Syncytial Virus (RSV)	
	□ 321 Rhinovirus (all species)	
	☐ 320 Rotavirus (all species)	
	□ 315 Rubella Virus	
	□ 302 Varicella Virus	
	☐ 348 West Nile Virus (WNV)	
508. Has	the recipient ever been infected with PCP / PJP?	

☐ Yes

CIBMTR Center Number: CIBMTR Research ID:			
□ No			
509. Does the recipient have GVHD due to maternal cell engraftment pre-HCT? (SCID only)			
□ Yes			
□ No			
Inherited Abnormalities of Platelets			
510. Specify inherited abnormalities of platelets classification			
☐ Congenital amegakaryocytosis / congenital thrombocytopenia (501)			
☐ Glanzmann thrombasthenia (502)			
☐ Other inherited platelet abnormality (509) – <i>Go to question 511</i>			
511. Specify other inherited platelet abnormality: – Go to end			
of form			
Inherited Disorders of Metabolism			
EAO Occasión imbanita della condensa of constable discondensa della cife estima			
512. Specify inherited disorders of metabolism classification			
☐ Osteopetrosis (malignant infantile osteopetrosis) (521)			
Leukodystrophies			
☐ Metachromatic leukodystrophy (MLD) (542)			
Adrenoleukodystrophy (ALD) (543) – <i>Go to question 514</i>			
☐ Krabbe disease (globoid leukodystrophy) (544)			
☐ Lesch-Nyhan (HGPRT deficiency) (522)			
□ Neuronal ceroid lipofuscinosis (Batten disease) (523)			
☐ Hereditary diffuse leukoencephalopathy with spheroids (HDLS) (551)			
Mucopolysaccharidoses ☐ Hurler syndrome (IH) (531)			
☐ Scheie syndrome (IS) (532)			
☐ Hunter syndrome (II) (533)			
☐ Sanfilippo (III) (534)			
☐ Morquio (IV) (535)			
☐ Maroteaux-Lamy (VI) (536)			
□ β-glucuronidase deficiency (VII) (537)			
☐ Mucopolysaccharidosis (V) (538)			
☐ Mucopolysaccharidosis, not otherwise specified (530)			

CIBMTR Center	Number: CIBMTR Research ID:	
Mu □	Colipidoses Gaucher disease (541)	
	Niemann-Pick disease (545)	
	I-cell disease (546)	
	Wolman disease (547)	
	Glucose storage disease (548)	
	Mucolipidoses, not otherwise specified (540)	
Po	lysaccharide hydrolase abnormalities Aspartyl glucosaminidase (561)	
	Fucosidosis (562)	
	Mannosidosis (563)	
	Polysaccharide hydrolase abnormality, not otherwise specified (560)	
	Other inherited metabolic disorder (529) – <i>Go to question 513</i>	
	Inherited metabolic disorder, not otherwise specified (520)	
	13. Specify other inherited metabolic disorder: G 14. Loes composite score: Adrenoleukodystrophy (ALD) only - Go to end of	Go to end of form
Histiocytic Dis	orders	
515. Sp	ecify histiocytic disorder classification	
Dis	seases of immune dysregulation, Familial Hemophagocytic Lymphohistiocytos Familial Hemophagocytic Lymphohistiocytosis, Perforin deficiency (FHL2) (2511)	sis (FHL)
	Familial Hemophagocytic Lymphohistiocytosis, UNC13D (FHL3) (2512)	
	Familial Hemophagocytic Lymphohistiocytosis, STX11 (FHL4) (2513)	
	Familial Hemophagocytic Lymphohistiocytosis, STXBP2 (FHL5) (2514)	
	Familial Hemophagocytic Lymphohistiocytosis, no mutation identified (2515)	
	Familial Hemophagocytic Lymphohistiocytosis, other mutations (2516)	
	Langerhans cell histiocytosis (histiocytosis-X) (572)	
	Hemophagocytosis (reactive or viral associated) (573)	
	Malignant histiocytosis (574)	
	Other histiocytic disorder (579) – Go to question 516	
	Histiocytic disorder, not otherwise specified (570)	
5	16. Specify other histiocytic disorder:	Go to end of

CIBMTR	R Ce	nter	Number:	CIBMTR Research ID:
5	517.	Did	the recipient	have an active or recent infection with a viral pathogen within 60 days of HCT?
			Yes – Go to	question 518
			No – <i>Go to</i>	question 519
		51	8. Specify v	riral pathogen <i>(check all that apply)</i>
			□ 304 A	denovirus
			□ 341 B	K Virus
			□ 344 C	Coronavirus
			□ 303 C	Cytomegalovirus (CMV)
			□ 347 C	hikungunya Virus
			□ 346 □	Pengue Virus
			□ 325 E	nterovirus (ECHO, Coxsackie)
			□ 327 E	interovirus D68 (EV-D68)
			□ 326 E	interovirus (polio)
			□ 328 E	Interovirus NOS
			□ 318 E	pstein-Barr Virus (EBV)
			□ 306 H	lepatitis A Virus
			□ 307 H	lepatitis B Virus
			□ 308 ⊢	lepatitis C Virus
			□ 340 H	lepatitis E
			□ 301 H	lerpes Simplex Virus (HSV)
			□ 317 H	luman herpesvirus 6 (HHV-6)
			□ 309 H	luman Immunodeficiency Virus 1 or 2
			□ 343 H	luman metapneumovirus
			□ 322 F	luman Papillomavirus (HPV)
			□ 349 H	luman T-lymphotropic Virus 1 or 2
			□ 310 Ir	nfluenza, NOS
			□ 323 Ir	nfluenza A Virus
			□ 324 Ir	nfluenza B Virus
			□ 342 J	C Virus (Progressive Multifocal Leukoencephalopathy (PML))
			□ 311 N	leasles Virus (Rubeola)
			□ 312 N	lumps Virus
			□ 345 N	lorovirus
			□ 316 H	luman Parainfluenza Virus (all species)
			□ 314 F	Respiratory Syncytial Virus (RSV)
			□ 321 F	thinovirus (all species)
			□ 320 F	otavirus (all species)

CIBMTR Cen	nter	Number: CIBMTR Research ID:
		□ 315 Rubella Virus
		□ 302 Varicella Virus
		□ 348 West Nile Virus (WNV)
	519	9. Has the recipient ever been infected with PCP / PJP
		☐ Yes – Go to end of form
		□ No – Go to end of form
Autoimmune	e Di	seases
520.	Spe	cify autoimmune disease classification
		nritis Rheumatoid arthritis (603)
		Psoriatic arthritis / psoriasis (604)
		Juvenile idiopathic arthritis (JIA): systemic (Still's disease) (640)
		Juvenile idiopathic arthritis (JIA): oligoarticular (641)
		Juvenile idiopathic arthritis (JIA): polyarticular (642)
		Juvenile idiopathic arthritis (JIA): other (643)
		Other arthritis (633)
		tiple sclerosis Multiple sclerosis (602)
		nective tissue diseases Systemic sclerosis (scleroderma) (607)
		Systemic lupus erythematosus (SLE) (605)
		Sjögren syndrome (608)
		Polymyositis / dermatomyositis (606)
		Antiphospholipid syndrome (614)
		Other connective tissue disease (634)
	Vas □	culitis Wegener granulomatosis (610)
		Classical polyarteritis nodosa (631)
		Microscopic polyarteritis nodosa (632)
		Churg-Strauss (635)
		Giant cell arteritis (636)
		Takayasu (637)
		Behcet syndrome (638)

CIBMTR Center	r Number: CIBM	R Research ID:
	Overlap necrotizing arteritis (639)	
	Other vasculitis (611)	
Oth	her neurological autoimmune diseases Myasthenia gravis (601)	
	Other autoimmune neurological disorder	(644)
Hei	ematological autoimmune diseases Idiopathic thrombocytopenic purpura (ITI	r) (645)
	Hemolytic anemia (646)	
	Evans syndrome (647)	
	Other autoimmune cytopenia (648) – Go	to question 521
Bo	owel diseases Crohn's disease (649)	
	Ulcerative colitis (650)	
	Other autoimmune bowel disorder (651)	- Go to question 522
Me □	etabolic Diabetes mellitus type 1 (660)	
	her Other autoimmune disease (629) – Go t	o question 523
52	21. Specify other autoimmune cytopenia:	
52	22. Specify other autoimmune bowel disc	rder:
52	23. Specify other autoimmune disease: _	Go to end of form
Tolerance Indu	uction Associated with Solid Organ Tran	splant
524. Spe	pecify solid organ transplanted (check all th	at apply)
	Kidney	
	Liver	
	Pancreas	
	Other organ – Go to question 525	
52	25. Specify other organ:	– Go to end of form

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526. Specify other disease:	– Go to end of form	