

Hematopoietic Cellular Transplant (HCT) Infusion

Registry Use Only
Sequence Number:
Date Received:

OMB No: 0915-0310 Expiration Date: 09/30/2028

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CIBM	ITR Center Number:		
CIBM	ITR Research ID:		
Even	t date:		·
	YYYY	MM	DD
HCT	type (check only one)		
	☐ Autologous		
Ī	☐ Allogeneic, unrelated		
	☐ Allogeneic, related		
Prod	uct type (check only one)		
	☐ Bone marrow		
	□PBSC		
	☐ Single cord blood unit		
	☐ Other product		
	Specify:		
NMD	P Product		
	□Yes		
	□ No		
Prod	uct Identifiers:		

CIBMTR Center Number: CIBM	TR Recipient ID:
NMDP cord blood unit ID:	NMDP donor ID:
Registry donor ID:	
Non-NMDP cord blood unit ID:	
Global Registration Identifier for Donors (GRID):	
Registry or UCB Bank ID:	
Donor DOB:	
Donor age:	old)
☐ Years	
Donor sex: ☐ Male ☐ Female	

CIBM	ITR Cente	r Number: CIBMTR Recipient ID:
A ser	ries of col	ne type of HCT product is infused, each product type must be analyzed and reported separately. Elections should be considered a <u>single product</u> when they are all from the same donor and use ection method and technique (and mobilization, if applicable), even if the collections are different days.
Pre-C	Collection	Therapy
1.	collection	lonor receive growth and mobilizing factors, prior to any stem cell harvest, to enhance the product of for this HCT? Allogeneic donors only — Go to question 2
	□ No –	Go to question 4
	2. Sp	ecify growth and mobilizing factor(s) <i>(check all that apply)</i> G-CSF (TBO-filgrastim, filgrastim, Granix, Neupogen and biosimilars) – <i>Go to question 4</i>
		Pegylated G-CSF (pegfilgrastim, Neulasta) – <i>Go to question 4</i>
		Motixafortide (Aphexda) – Go to question 4
		Plerixafor (Mozobil and biosimilars) – <i>Go to question 4</i>
		Other growth or mobilizing factor(s) – <i>Go to question 3</i>
	3.	Specify other growth or mobilizing factor(s):
Prod	uct Collec	ction
4.	Date of f	irst collection for this mobilization:
5.		ticoagulants or other agents added to the product between collection and infusion? – <i>Go to question 6</i>
	□ No -	Go to question 8
	6. Sp	pecify anticoagulant(s) or other agents <i>(check all that apply)</i> Acid citrate dextrose (ACD, ACD-A) – <i>Go to question 8</i>
		Citrate phosphate dextrose (CPD, CPD-A) – Go to question 8
		Dimethylsulfoxide (DMSO) – Go to question 8
		Ethylenediaminetetraacetic acid (EDTA) – <i>Go to question 8</i>
		Heparin– Go to question 8
		Plasmalyte– Go to question 8
		Other agent – Go to question 7
	7	. Specify other agent:

CIBM	TR Ce	enter Number: CIBMTR Recipient ID:
Prod	uct Tra	ansport and Receipt
8.		this product collected off-site and shipped to your facility? ′es – <i>Go to question</i> 9
		lo – Go to question 22
	9.	Date of receipt of product at your facility:
	10.	Time of receipt of product (24-hour clock): : : □ standard time □ daylight savings time
	11.	Specify the shipping environment of the product(s) ☐ Room temperature – <i>Go to question 13</i>
		□ Cooled (refrigerator temperature, not frozen, refrigerated gel packs or Credo Cube TM transporter) – Go to question 13
		☐ Frozen (cryopreserved) – Go to question 13
		☐ Other shipping environment – Go to question 12
		12. Specify other shipping environment:
	13.	Was there any indication that the environment within the shipper was outside the expected temperature range for this product at any time during shipment? ☐ Yes
		□ No
	14.	Were the secondary containers (e.g., insulated shipping containers and unit cassette) intact when they arrived at your center? ☐ Yes
		□ No
	15.	Was the cord blood unit stored at your center prior to thawing? (Cord blood units only) ☐ Yes – Go to question 16
		□ No – Go to question 19
		16. Specify the storage method used for the cord blood unit ☐ Electric freezer
		□ Liquid nitrogen
		□ Vapor phase

CIBM	ITR Ce	nter Number: CIBMTR Recipient ID:	CIBMTR Recipient ID:			
		17. Temperature during storage □ ≤-150° C				
		□ -149° C to ≥ -80° C				
		18. Date storage started:				
		total number of cells (not cells per kilogram) prior to cryopreservation: (Information provided for t cord blood bank).	he			
	19.	Total nucleated cells: • x 10 (Includes nucleated red and nucleated white cells) (Cord blood units only)				
	20.	CD34+ cells (Cord blood units only) □ Done – Go to question 21				
		□ Not done – Go to question 22				
		21. Total number of CD34+ cells: • x 10				
Prod	uct Pr	ocessing / Manipulation				
22.		he product thawed from a cryopreserved state prior to infusion? es – <i>Go to question 23</i>				
	□ N	o – Go to question 30				
	23.	Was the entire product received by the center thawed? ☐ Yes				
		□ No				
	24.	Date thawing process initiated:				
	25.	Time at initiation of thaw (24-hour clock): :				
	26.	Time of thaw completion (24-hour clock): : □ standard time □ daylight savings time				
	27.	What method was used to thaw the product? ☐ Water bath – Go to question 29				
		□ Electric warmer – <i>Go to question 29</i>				
		□ Other method – <i>Go to question 28</i>				

CIBN	ITR C	enter	Num	ber: CIBMTR Recipient ID:
		28.	Spe	cify other method used to thaw the product:
	29.		any Yes	incidents or product complaints occur while preparing or thawing the product?
			No	
30.		•		ct processed prior to infusion? to question 31
		No –	Go to	o question 32
	31.	Spe	•	processing (check all that apply) y coat enriched (buffy coat preparation)
			Dilu	ted
			Plas	ma reduced
			RBC	Creduced
			Was	shed
32.		-		ct manipulated prior to infusion? to question 33
		No –	Go to	o question 42
	33.	Spe	-	manipulations performed <i>(check all that apply)</i> vivo expansion – Go to question 34
			Gen	etic manipulation (gene transfer / transduction) – <i>Go to question</i> 36
			CD3	4 enriched (CD34+ selection) – Go to question 42
			Ex-\	rivo T-cell depletion – <i>Go to question</i> 37
			Neg	ative fraction – <i>Go to question 42</i>
			Othe	er manipulation – Go to question 41
		34.	Spe	cify ex-vivo expansion Omidubicel (OMISIRGE)
				Other method
			35.	Specify other ex-vivo expansion:
		36.	Spe	cify genetic manipulation: (gene transfer / transduction)
		37.	Spe	cify antibodies used <i>(check all that apply)</i> Anti CD3
			П	Anti CD4

CIBMTR Cente	er Numl	ber: CIBMTR Recipient ID:
		Anti CD8
		Anti CD19
		Anti CD45RA
		α/β Antibody
		Anti CD52
		Other antibody– <i>Go to question</i> 38
	38.	Specify other antibody used:
39). Sped	cify T-cell depletion method Antibody affinity column
		Immunomagnetic beads
		Other method – Go to question 40
	40.	Specify other T-cell depletion method:
41	. Spec	cify other cell manipulation:
Product Analy	sis at	Infusion
42. Date of բ	oroduct	analysis:
43. Total vol	ume of	f product received by center plus additives: •mL
In this section	, repo	rt the total number of cells (not cells per kilogram) and do not correct for viability.
		cells (TNC) (whole product) to question 45
□ Not	done –	Go to question 50
45. To	otal nuc	cleated cells: • x 10
46. Vi □	-	of TNC e – Go to question 47
	Not	done – Go to question 50
47.	Viab	ility of TNC: %

CIBMTR Cer		enter	Num	ber: CIBMTR Recipient ID:
	4	l8.	Meth □	nod of testing TNC viability Flow cytometry based (includes 7-AAD, AOPI, and AOEB) – Go to question 50
				Trypan blue – Go to question 50
				Other method – Go to question 49
			49.	Specify other method of testing TNC viability:
50.				e blood cells to question 51
		Not do	one –	Go to question 52
	51.	Tota	al nur	mber of nucleated white blood cells: • x 10
52.	Mond			ells o to question 53
		Not do	one –	Go to question 54
	53.	Tota	al nur	mber of mononuclear cells: • x 10
54.				blood cells to question 55
		Not do	one –	Go to question 56
	55.	Tota	al nur	mber of nucleated red blood cells: • x 10
56.	CD34			to question 57
	□ 1	Not do	one –	Go to question 62
	57.	Tota	al nur	mber of CD34+ cells: • x 10
	58.	Vial □	-	of CD34+ cells e – Go to question 59
			Not	done – Go to question 62
		59.	Viab	ility of CD34+ cells: %
		60.	Meth □	nod of testing CD34+ cell viability Flow cytometry based (7-AAD, AOPI, and AOEB) – Go to question 62

CIBM	ITR C	enter	Num	ber: CIBMTR Recipient ID:
				Trypan blue – Go to question 62
				Other method – Go to question 61
			61.	Specify other method of testing CD34+ cell viability:
62.		+ cell		to question 63
				Go to question 68
	63.	Tot	al nui	mber of CD3+ cells: • x 10
	64.	Via □	-	of CD3+ cells e – Go to question 65
			Not	done – Go to question 68
		65.	Viab	ility of CD3+ cells:
		66.	MetI □	nod of testing CD3+ cell viability Flow cytometry based (7-AAD, AOPI, and AOEB) – Go to question 68
				Trypan blue- Go to question 68
				Other method – Go to question 67
			67.	Specify other method of testing CD3+ cell viability:
68.			l+ ce – G c	ls o to question 69
		Not d	one -	Go to question 70
	69.	Tot	al nu	mber of CD3+CD4+ cells: • x 10
70.			3+ ce - G c	ls o to question 71
		Not d	one -	Go to question 72
	71.	Tot	al nu	mber of CD3+CD8+ cells: • x 10
72.				y-forming units (CFU) assessed after thawing? (Cord blood units only) to question 73
		No –	Go to	question 78
	73.	Wa	s the	re growth?

CIBN	ITR (Center	Number: CIBMTR Recipient ID:
			Yes
		_	No
	74.	Ind	licate which assessments were carried out <i>(check all that apply)</i> Total CFU-GM – <i>Go to question 75</i>
			Total CFU-GEMM – <i>Go to question 76</i>
			Total BFU-E – <i>Go to question 77</i>
		75.	. Total CFU-GM: • x 10
		76.	. Total CFU-GEMM: • x 10
		77.	. Total BFU-E: • x 10
78.	(co	mplete	positive cultures (for bacterial or fungal infections) obtained from the product at the transplant center? e for all cell products) - Go to question 79
		No –	Go to question 84
		Pend	ling - Go to question 84
		Unkn	nown– Go to question 84
	Sp	ecify o	organism code(s):
	79.		
	80.		
	81.		
	82.		
		83.	Specify other organism:
			odes for Commonly Reported Organisms Bacterial Infections
			121 Acinetobacter (all species)
			125 Bordetella pertussis (whooping cough)
			128 Campylobacter (all species)
			129 Capnocytophaga (all species)
			171 Chlamydia (pneumoniae)

CIBMTR Center	Number: CIBMTR Recipient ID:
	130 Citrobacter (freundii, other species)
	131 Clostridium (all species except difficile)
	132 Clostridium difficile
	173 Corynebacterium jeikeium
	196 Cutibacterium acnes (Propionibacterium)
	134 Enterobacter (all species)
	135 Enterococcus (all species)
	177 Enterococcus, vancomycin resistant (VRE)
	136 Escherichia (also E. coli)
	139 Fusobacterium (all species)
	187 Haemophilus influenzae
	188 Haemophilus non-influenzae
	146 Klebsiella (all species)
	147 Lactobacillus (bulgaricus, acidophilus, other species)
	189 Legionella pneumophila
	190 Legionella non-pneumophila
	103 Leptospira (all species)
	148 Leptotrichia buccalis
	149 Leuconostoc (all species)
	104 Listeria monocytogenes
	151 Micrococcus, NOS
	118 Mycobacterium abscessus
	112 Mycobacterium avium - intracellulare (MAC, MAI)
	108 Mycobacterium cheloneae
	109 Mycobacterium fortuitum
	114 Mycobacterium haemophilum
	115 Mycobacterium kansasii
	116 Mycobacterium marinum
	117 Mycobacterium mucogenicum
	110 Mycobacterium tuberculosis (tuberculosis, Koch bacillus)
	105 Mycoplasma (all species)
	183 Neisseria gonorrhoeae
	184 Neisseria meningitidis
	106 Nocardia (all species)
П	153 Pastouralla multocida

CIBMTR Center	Number: CIBMTR Recipient ID:
	155 Proteus (all species)
	157 Pseudomonas or Burkholderia cepacia
	185 Pseudomonas aeruginosa
	186 Pseudomonas non-aeruginosa
	159 Rhodococcus (all species)
	107 Rickettsia (all species)
	160 Salmonella (all species)
	161 Serratia marcescens
	162 Shigella (all species)
	180 Staphylococcus aureus (Methicillin Resistant)
	179 Staphylococcus aureus (Methicillin Sensitive)
	158 Stenotrophomonas maltophilia
	166 Stomatococcus mucilaginosis
	181 Streptococcus, alpha-hemolytic
	182 Streptococcus, Group B
	178 Streptococcus pneumoniae
	168 Treponema (syphilis)
	169 Vibrio (all species)
Fun	ngal Infections
	210 Aspergillus, NOS
	211 Aspergillus flavus
	212 Aspergillus fumigatus
	213 Aspergillus niger
	215 Aspergillus terreus
	214 Aspergillus ustus
	270 Blastomyces (dermatitidis)
	201 Candida albicans
	208 Candida non-albicans
	271 Coccidioides (all species)
	222 Cryptococcus gattii
	221 Cryptococcus neoformans
	230 Fusarium (all species)
	261 Histoplasma (capsulatum)
	241 Mucorales (all species)
	260 Pneumocystis (PCP / PJP)

CIBMTR Center Number:		IBMTR Recipient ID: _		
		242 Rhizopus (all species)		
		272 Scedosporium (all species)		
		240 Zygomycetes, NOS		
		503 Suspected fungal infection		
		777 Other organism		
Prod	uct Infusi	on		
84.	Date of the	nis product infusion:		
85.		entire volume of the product received - Go to question 88	by the center infused?	
	□ No –	Go to question 86		
	86. Sp	ecify what happened to the reserved posterior in the control of th	portion (check all that a	ppply)
		Cryopreserved for future use – Go t	o question 88	
		Research - Go to question 88		
		Training or Quality Control – Go to	question 88	
		Other fate – Go to question 87		
	87	. Specify other fate:		-
88.	Time pro	duct infusion initiated (24-hour clock):	Hour • Minute	☐ standard time ☐ daylight savings time
89.	Date infu	sion stopped:		
90.	Time pro	duct infusion completed (24-hour cloc	k): :Minute	☐ standard time ☐ daylight savings time
91.		ne route of product infusion venous – Go to question 93		
	□ Intra	medullary (<i>Intraosseous</i>) – Go to que	stion 93	
	□ Othe	r route of infusion – Go to question 9	02	
	92. Sp	ecify other route of infusion:		

CIBMTR Center Number:		enter Number:	CIBMTR Recipient ID:		
		ring questions are applicable to 34. Autologous and NMDP prod	cord blood units only. Non-NMDP allogeneic products continue with ucts go to end of form.		
93.		e there any adverse events or incid Yes – Go to question 94	dents associated with the stem cell infusion?		
	□ 1	No – Go to question 134			
	Spec	cify the following adverse event	(s)		
	94.	Brachycardia □ Yes – Go to question 95			
		□ No – Go to question 96			
		95. In the clinician's judgment, v ☐ Yes	vas the adverse event a direct result of the infusion?		
		□ No			
	96.	Chest tightness / pain ☐ Yes – Go to question 97			
		□ No – Go to question 98			
		97. In the clinician's judgment, v □ Yes	as the adverse event a direct result of the infusion?		
		□ No			
	98.	Chills at time of infusion ☐ Yes – <i>Go to question 99</i>			
		□ No – Go to question 100			
		99. In the clinician's judgment, v □ Yes	as the adverse event a direct result of the infusion?		
		□ No			
	100.	Fever ≤ 103° F within 24 hours o	f infusion		
		□ No – Go to question 102			
		101. In the clinician's judgment ☐ Yes	t, was the adverse event a direct result of the infusion?		
		□ No			

CIBMTR Cer	nter Number: CIBMTR Recipient ID:
	□ Yes – Go to question 103
	□ No – Go to question 104
	103. In the clinician's judgment, was the adverse event a direct result of the infusion?☐ Yes
	□ No
	Gross hemoglobinuria □ Yes – <i>Go to question 105</i>
	□ No – Go to question 106
	105. In the clinician's judgment, was the adverse event a direct result of the infusion?☐ Yes
	□ No
	Headache □ Yes– Go to question 107
	□ No – Go to question 108
	107. In the clinician's judgment, was the adverse event a direct result of the infusion?☐ Yes
	□ No
108.	Hives □ Yes – Go to question 109
	□ No – Go to question 110
	109. In the clinician's judgment, was the adverse event a direct result of the infusion?☐ Yes
	□ No
	Hypertension □ Yes – <i>Go to question 111</i>
	□ No – Go to question 112
	111. In the clinician's judgment, was the adverse event a direct result of the infusion?☐ Yes
	□ No

112. Hypotension

CIBMTR Center Nu	mber: CIBMTR Recipient ID:
□ Ye	es – Go to question 113
□ No	o – Go to question 114
	In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
	□ No
	ia requiring oxygen (O₂) support es – Go to question 115
□ No	o – Go to question 116
	In the clinician's judgment, was the adverse event a direct result of the infusion? □ Yes
	□ No
116. Nause □ Ye	a es – Go to question 117
□ No	o – Go to question 118
117.	In the clinician's judgment, was the adverse event a direct result of the infusion? □Yes
	□No
118. Rigors □ Ye	, mild es – Go to question 119
□ No	o – Go to question 120
	In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
	□ No
120. Rigors □ Ye	, severe es – Go to question 121
□ No	o – Go to question 122
	In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
	□ No

122. Shortness of breath (SOB)

CIBMTR Center	Number: CIBMTR Recipient ID:
	Yes – Go to question 123
	No – Go to question 124
123.	In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
	□ No
124. Tad	chycardia Yes – Go to question 125
	No – Go to question 126
125	5. In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
	□ No
126. Vor □	miting Yes – Go to question 127
	No – Go to question 128
127	7. In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
	□ No
	ner expected AE Yes – <i>Go to question 129</i>
	No – Go to question 131
129	D. Specify other expected AE:
130	In the clinician's judgment, was the adverse event a direct result of the infusion?☐ Yes
	□ No
	ner unexpected AE Yes – Go to question 132
	No – Go to question 134
132	2. Specify other unexpected AE:

CIBMTR Center Number:		lumber: CIBMTR Recipient ID:
	133.	In the clinician's judgment, was the adverse event a direct result of the infusion? ☐ Yes
		□ No
Dono	r / Infant De	mographic Information
alloge		ographic Information section (questions 134-159) is to be completed for all non-NMDP s. If the stem cell product was from an NMDP donor or an autologous donor, continue to the
134.		nor ever pregnant? Go to question 135
	□ No – G	o to question 137
	□ Unknov	vn – Go to question 137
		ber of pregnancies Known – <i>Go to question 136</i>
		Jnknown – Go to question 137
	136.	Specify number of pregnancies:
137.		ancestry (select one or more options that closest identifies the donor's background) - Go to question 138
	□ Black o	r African – Go to question 138
	□ Hispani	ic or Latino – <i>Go to question 138</i>
	□ Indigen	ous American – Go to question 138
	☐ Jewish	– Go to question 138
	□ Middle	Eastern or North African – <i>Go to question 138</i>
	□ Pacific	Islander – Go to question 138
	□ White -	Go to question 138
	□ Not oth	erwise specified – Go to question 139
	□ Prefer r	not to answer – <i>Go to question 139</i>
	138. Geog	graphic ancestry detail (select one or more options that closest identifies the donor's background)
	Asia	an
		Caribbean Indian
		Chinese
	□ F	Filipino

CIBMTR Center N	Number: CIBMTR Recipient ID:
	Indian
	Japanese
	Korean
	Malaysian
	Mongolian
	Pakistani
□ .	Taiwanese
	Thai
– '	Vietnamese
	Other Indian Subcontinent (e.g. Bangladeshi, Nepali, Sri Lankan, etc.)
	Other Southeast Asian (e.g. Cambodian, Indonesian, Singaporean, etc.)
	Not otherwise specified Asian
Bla	ck or African
	African American
	Black Caribbean (e.g. Haitian, Jamaican, etc.)
	Black South or Central American
_	East African (e.g. Ethiopian, Kenyan, Somali, Tanzanian, etc.)
_	South African (e.g. Angolan, Botswanan, Mozambican, Zambian, Zimbabwean, etc.)
	West African (e.g. Ghanian, Malian, Nigerian, Senegalese, etc.)
	Not otherwise specified Black/African
	·
His	panic or Latino
	Brazilian
	Caribbean Hispanic (e.g. Dominican)
	Cuban
	Mexican
	Puerto Rican
	South / Central American Hispanic
	Not otherwise specified Hispanic / Latino
Indi	igenous American
	Alaska Native
	Indigenous Caribbean
	Indigenous North American
	Indigenous South / Central American

CIBMTR Center Number:		Number: CIBMTR Recipient ID:
		Not otherwise specified Indigenous American
		e <mark>wish</mark> Ashkenazi
		Mizrahi
		Sephardi
		Not otherwise specified Jewish
		iddle Eastern or North African Arab Peninsula (e.g. Emirati, Kuwaiti, Saudi, Yemeni etc.)
		Central Asian (e.g. Afghan, Iranian, Kazakhstani, Turkish, etc.)
		East Mediterranean (e.g. Iraqi, Jordanian, Lebanese, Syrian, etc.)
		North African (e.g. Algerian, Egyptian, Moroccan, etc.)
		Not otherwise specified Middle Eastern / North African
		acific Islander Melanesian (e.g. Fijian, Papua New Guinean, etc.)
		Micronesian (e.g. Chamorro, Guamanian, Marshallese, etc.)
		Native Hawaiian
		Polynesian (e.g. Māori, Samoan, Tongan, etc.)
		Not otherwise specified Pacific Islander
	W	hite
		Eastern European (e.g. Bulgarian, Georgian, Polish, Romanian, Ukrainian etc.)
		Northern European (e.g. Finnish, Norwegian, Swedish etc.)
		Russian or Former Soviet Union
		Southern European (e.g. Greek, Italian, Portuguese, Spanish, etc.)
		Western European (e.g. British, French, German, Irish, Scottish, etc.)
		White Caribbean
		White South or Central American
		Not otherwise specified White
		donor a carrier for potentially transferable genetic diseases? Go to question 140
	□ No- (Go to question 142
	140. Sp€	ecify potentially transferable genetic disease (check all that apply) Sickle cell anemia
		Thalassemia

CIBMTE	R Ce	nter	Number: CIBMTR Recipient ID:
			Other hemoglobinopathy
			Other disease– Go to question 141
	1	41.	Specify other transferable genetic disease:
142. V			lonor / product tested for other transferable genetic or clonal abnormalities? Go to question 143
	J N	lo –	If this is a related donor, go to question 148; all other donor types go to end of form
	J U	nkn	own – If this is a related donor, go to question 148; all other donor types go to end of form
1	43.		nal hematopoiesis of indeterminate potential (CHIP) Yes– Go to question 144
			No- Go to question 145
		144	. What was the method of testing used?
1	45.		noclonal B-cell lymphocytosis Yes
			No
1	46.		er transferable genetic or clonal abnormality Yes– <i>Go to question 147</i>
			No- Go to question 148
		147	7. Specify other transferable genetic or clonal abnormality:
	ologo	ous	uestions (148 - 159) apply only to allogeneic related donors. If the stem cell product was from donor, Non-NMDP unrelated donor, NMDP donor, or was a cord blood unit, then continue to the .
148. D		is do	onor have a central line placed?
	J N	lo	
149. V		he d	lonor hospitalized (inpatient) during or after the collection?
	J N	lo	
150. D			onor experience any life-threatening complications during or after the collection? Go to question 151
	J N	lo –	Go to question 152

CIBM	ITR Center Number:
	151. Specify life-threatening complication experienced:
152.	Did the allogeneic donor give one or more autologous transfusion units? ☐ Yes – <i>Go to question 153</i>
	□ No – Go to question 155
	153. Date of collection: DD
	154. Number of units:
155.	Did the donor receive blood transfusions as a result of the collection? <i>(check all that apply)</i> ☐ Autologous transfusions – <i>Go to question 156</i>
	☐ Allogeneic transfusions— Go to question 157
	□ No – Go to question 158
	156. Specify number of autologous units:
	157. Specify number of allogeneic units:
158.	Did the donor die as a result of the collection? ☐ Yes – Go to question 159
	□ No – Go to end of form
	159. Specify cause of death: